

Positive selection on non-coding sequences during primate evolution: from genome to nucleotide

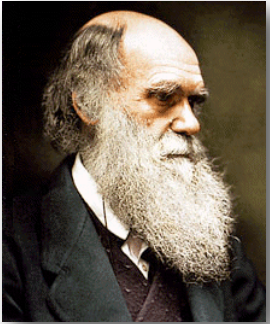
Gregory Wray

**Department of Biology
and
Institute for Genome Sciences & Policy**

**Duke University
Durham, NC, USA**



The great apes and an outgroup



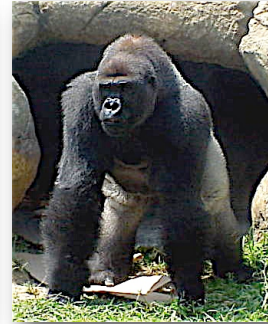
Homo sapiens



Pan troglodytes



Pan paniscus



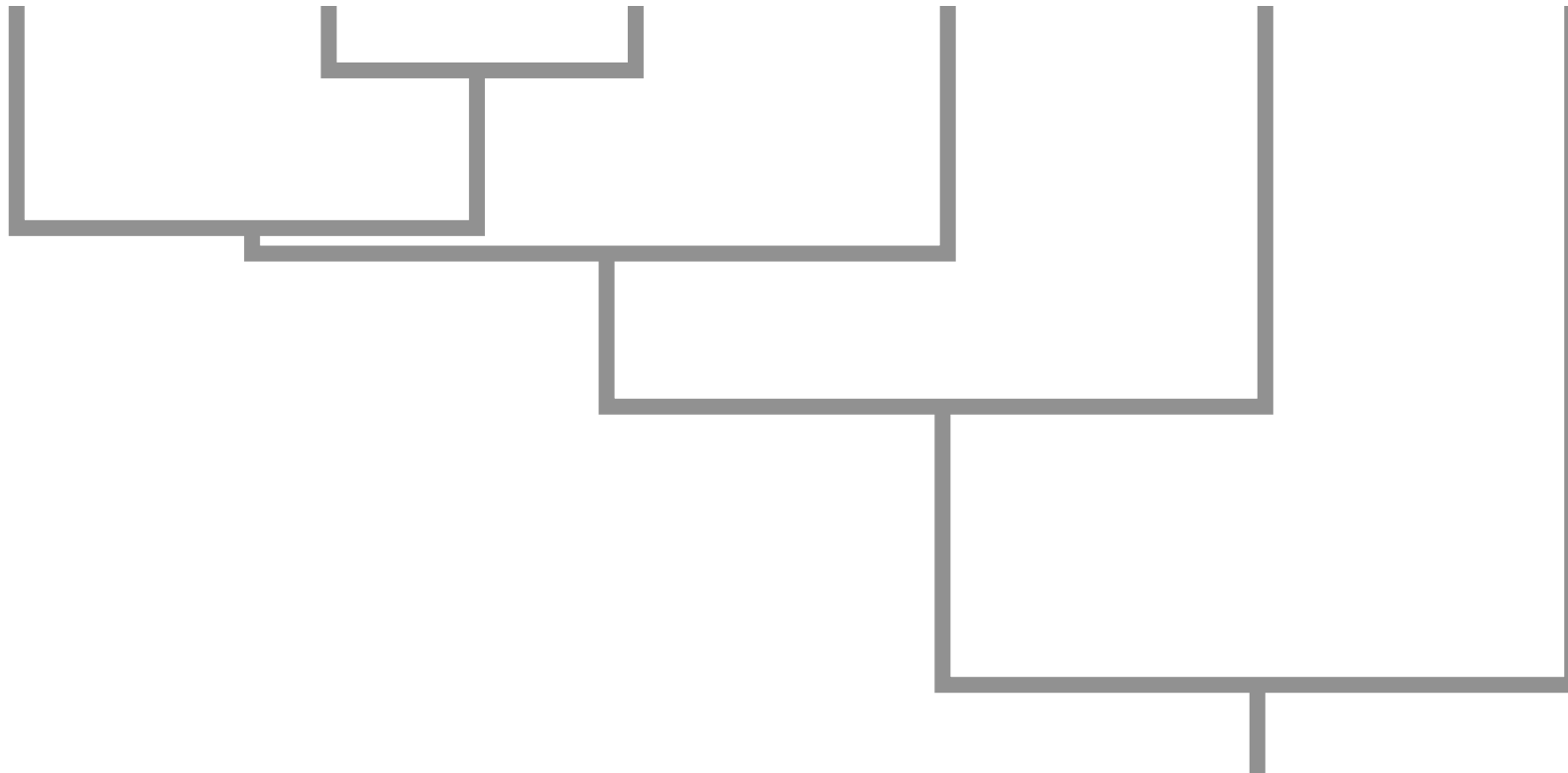
Gorilla gorilla



Pongo pygmaeus



Macaca mulatta



An apparent paradox

Genotype: quite similar

Very similar gene complement

Many identical proteins

~98.8% sequence identity

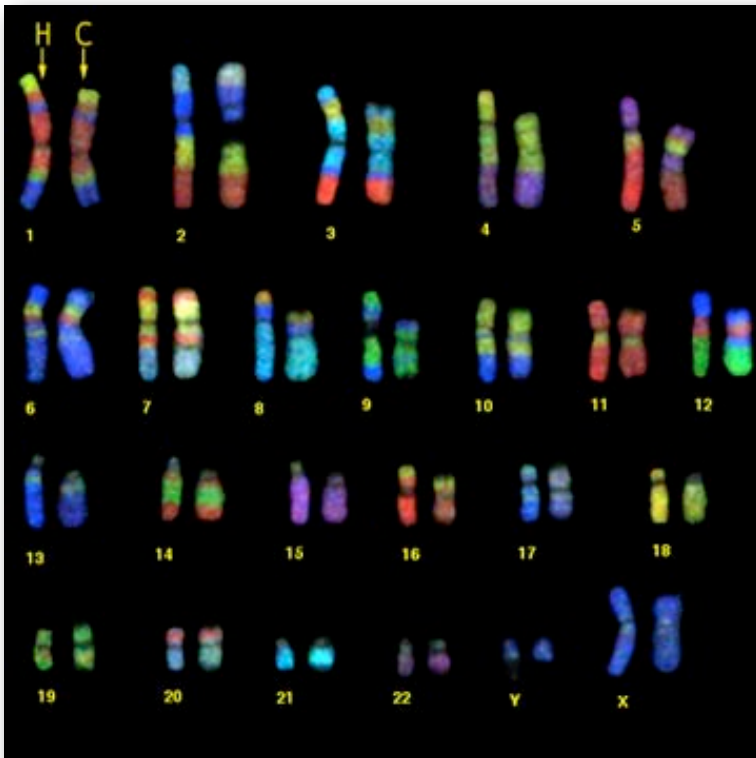
Phenotype: many differences

Cognition and behavior

Morphology

Physiology and reproduction

Disease susceptibility



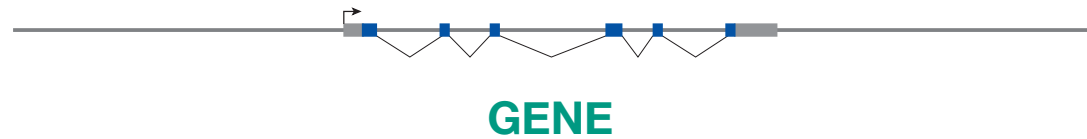
Michael Neugebauer

King and Wilson's proposal

Phenotypic differences between humans and chimpanzees are due primarily to changes in the regulation of gene expression

King, M-C and A Wilson (1975) *Science* 188:107

The problem: finding a needle in a field of haystacks



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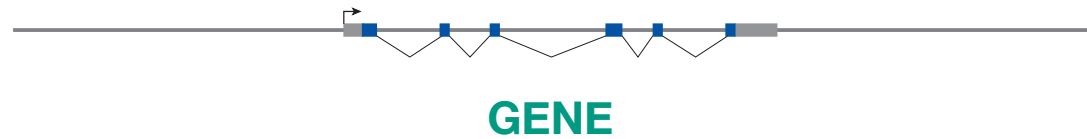
MUTATION

The problem: finding a needle in a field of haystacks

~24,000 genes



1 gene



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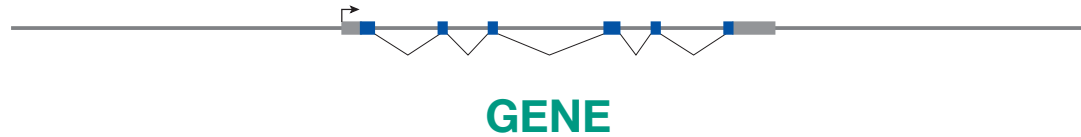


1 gene

10,000's of nucleotides



1 nucleotide



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MUTATION

Finding the needles

Candidate gene approach:

Think of genes that seem plausibly associated with trait of interest

Figure out a way to test for functional & phenotypic consequences

Often works, but limited to a few, well-studied genes

Finding the needles

Candidate gene approach:

Think of genes that seem plausibly associated with trait of interest
Figure out a way to test for functional & phenotypic consequences
Often works, but limited to a few, well-studied genes

Open-ended screen:

Identify genes that show evidence of recent positive selection
Figure out a way to test for functional & phenotypic consequences
Unbiased, considers all genes (even if no functional data)

screening for more candidate genes

testing for functional differences

connecting the dots: 3 case studies

screening for more candidate genes

testing for functional differences

connecting the dots: 3 case studies

A scan for positive selection on non-coding sequences



Goals: unbiased, genome-wide, statistically robust

Three-genome comparison

Human, chimpanzee, macaque

Scan non-coding regions near as many genes as possible



Test for selection using a random-effects likelihood approach

Use introns as local neutral proxies

Estimate substitution rates under different models

Use likelihood ratio to test for significance



Haygood et al. (2007) *Nature Genetics* 39:1140-1144.

Testing for positive selection in non-coding sequences



$$\zeta = \frac{\text{query}}{\text{reference}} = \frac{\text{red bar}}{\text{green bar}}$$

$$\zeta < 1$$

negative selection
functional constraint

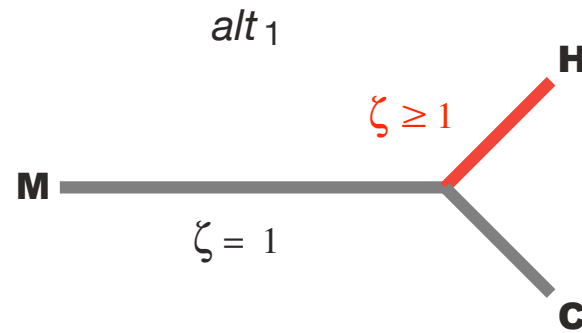
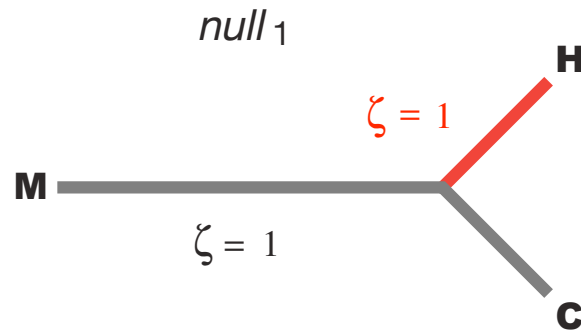
$$\zeta = 1$$

drift

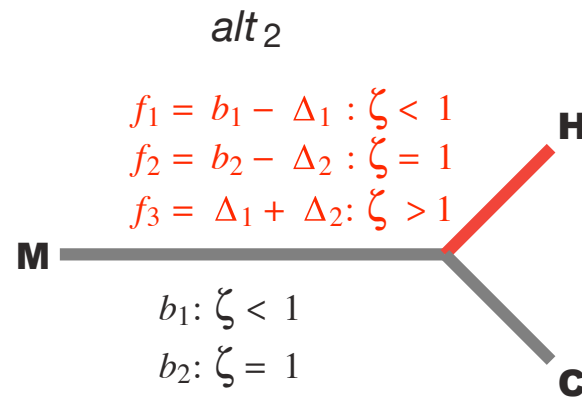
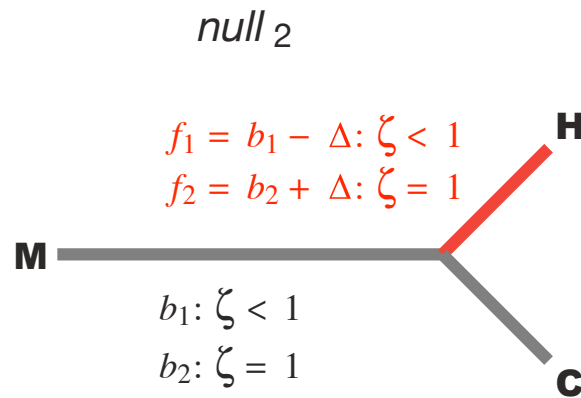
$$\zeta > 1$$

positive selection
adaptation

Likelihood-based tests for positive selection

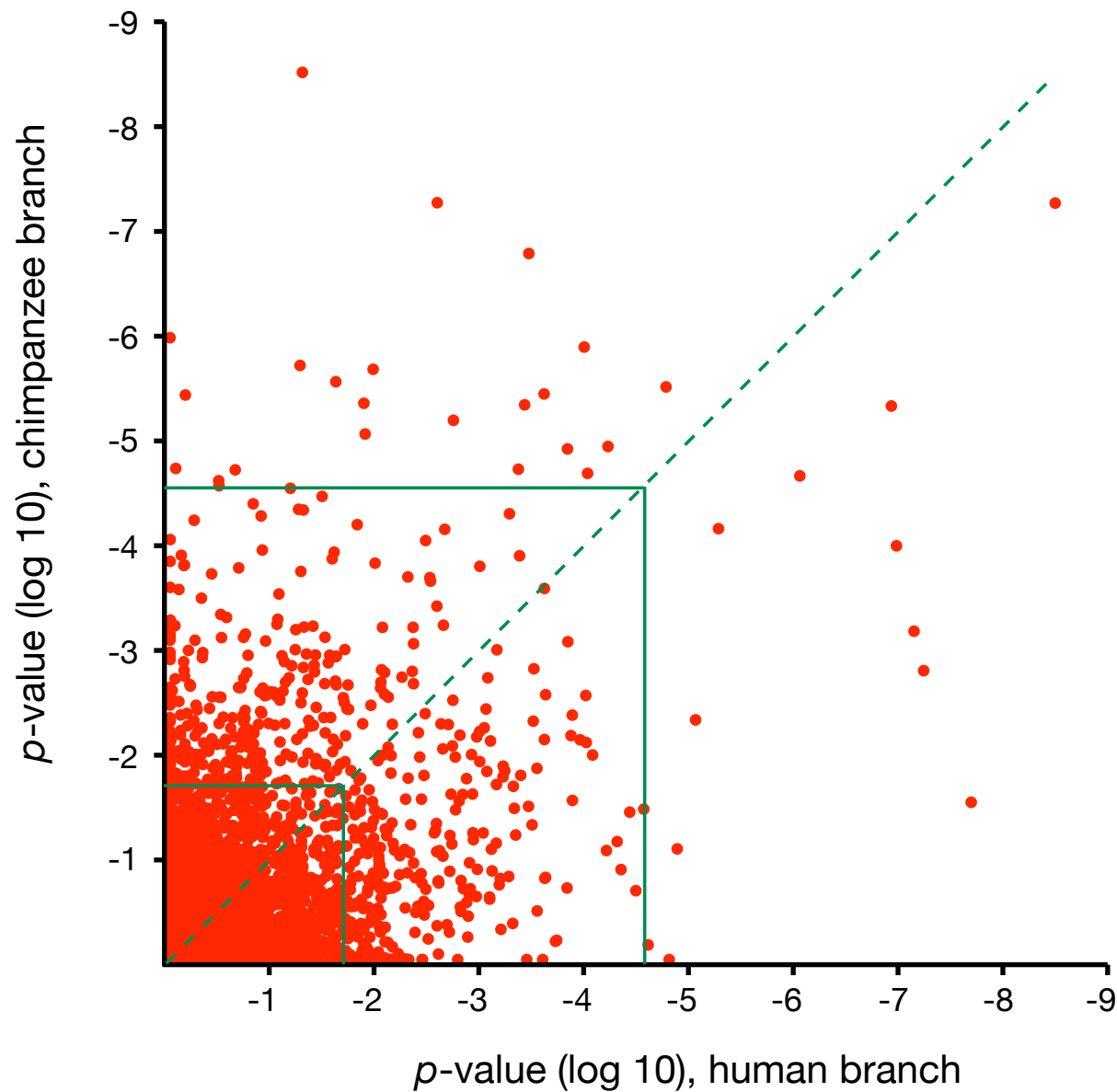


fewer assumptions,
unrealistic
less sensitive

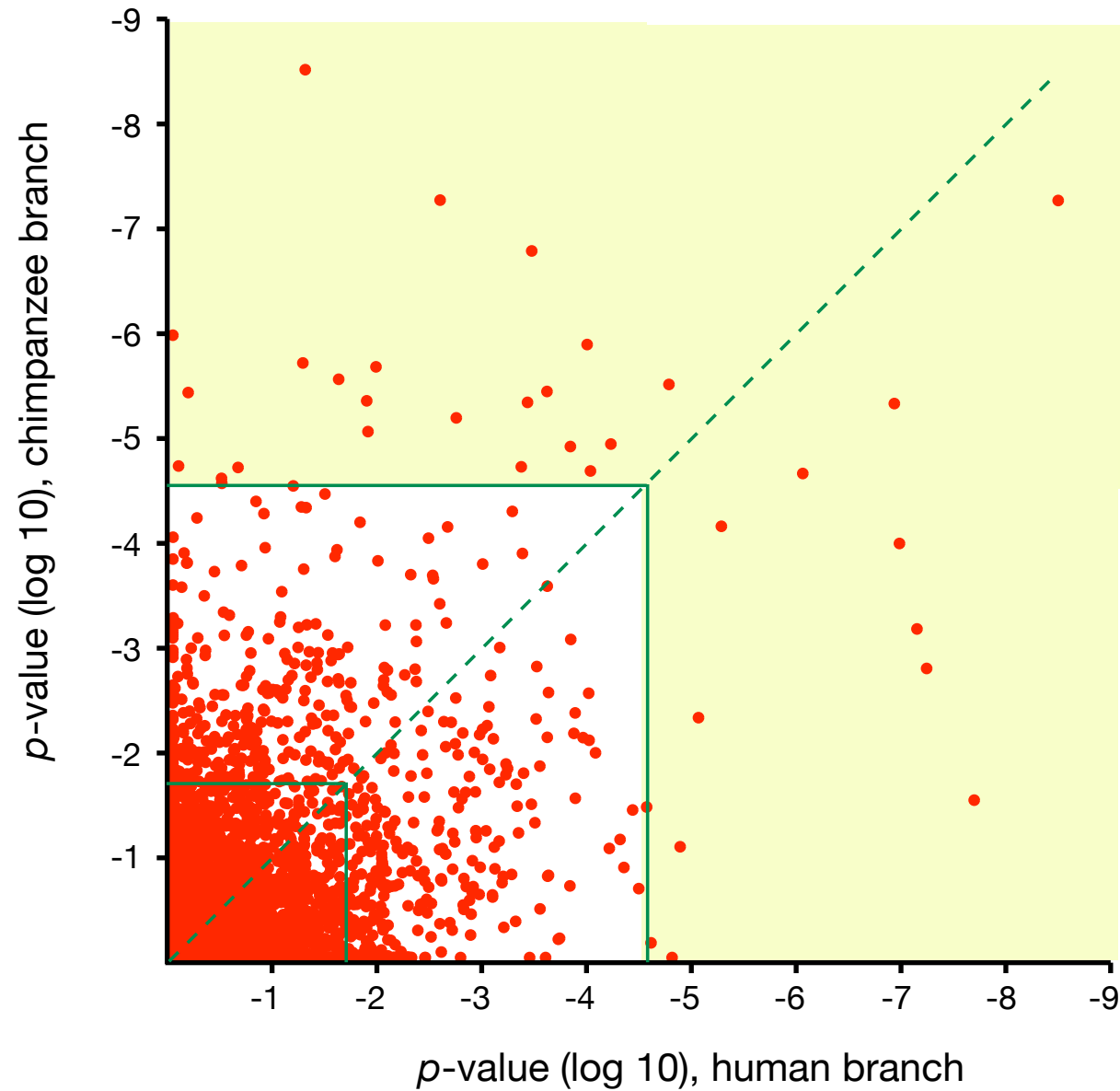


more realistic,
more sensitive,
harder to estimate

Distribution of p -values on chimp and human branches



Distribution of p -values on chimp and human branches



No signal of adaptation in neural function from coding regions

Inferring Nonneutral Evolution from Human-Chimp-Mouse Orthologous Gene Trios

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gence caused by random drift and divergence driven by natural selection. Early observations of unexpectedly low levels of protein divergence between humans and chimpanzees led to the hypothesis that most of the evolutionary changes must have occurred at the level of gene regulation (*1*). Recently, much more extensive efforts at DNA sequencing in nonhuman primates has confirmed the very close evolutionary relationship between humans and chimps (*2*), with an

average nucleotide divergence of just 1.2% (*3–5*). The role of protein divergence in causing morphological, physiological, and behavioral differences between these two species, however, remains unknown.

Here we apply evolutionary tests to identify genes and pathways from a new collection of more than 200,000 chimpanzee exonic sequences that show patterns of divergence consistent with natural selection along the human and chimpanzee lineages.

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^d Beijing Genomics Institute, Chinese Academy of Sciences, Beijing, China

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^f Department of Biochemistry and Molecular Biology, University of Southern Denmark, DK-5230, Odense M, Denmark

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No enrichment for brain-expressed genes
No enrichment for neural developmental or neural function

Over-represented functional categories: 5' flanking region

category	# genes	<i>p</i> -value on:	
		human branch	chimp branch
protein folding	70	0.0067	0.77
other neuronal activity	31	0.013	0.039
neurogenesis	133	0.013	0.032
glycolysis	21	0.014	0.72
carbohydrate metabolism	210	0.020	0.017
ectoderm development	169	0.020	0.11
mesoderm development	161	0.024	0.17
synaptic transmission	25	0.025	0.34
vision	64	0.025	0.15
oncogene	25	0.045	0.46
anion transport	31	0.049	0.17

Human branch, *alt2-null2* model contrast. Biological Process categories based on Panther Classification System (www.pantherdb.org). *p*-value = one-tailed Mann-Whitney test for human or chimpanzee branch. *p*-value < 0.01, *p*-value < 0.03, *p*-value < 0.03

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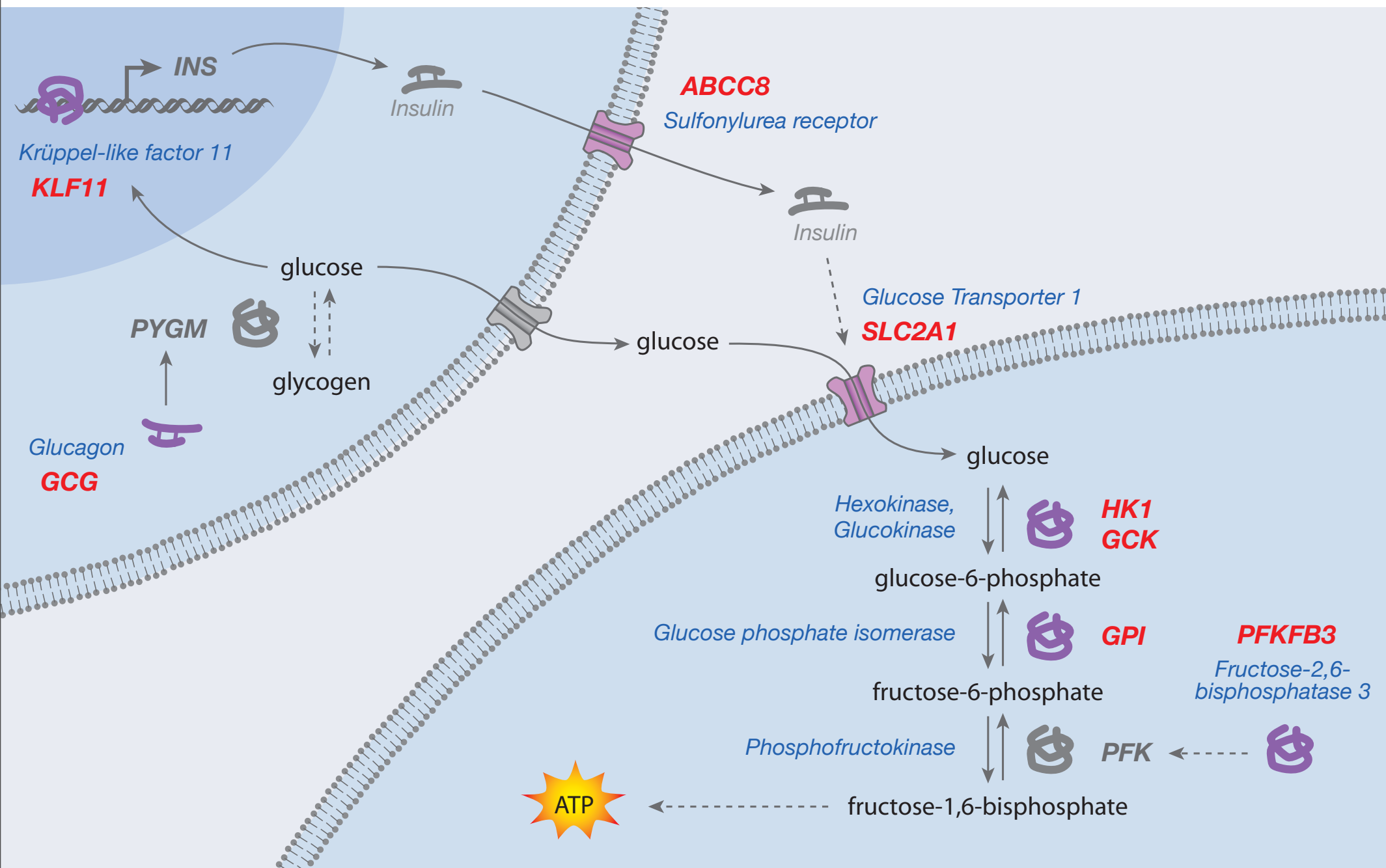
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Positive selection on a functional cluster: sugar metabolism



Why positive selection on regulation of sugar metabolism?

Shift in diet

Humans are an outlier among the great apes in terms of diet

Other great apes consume primarily fruits, nuts, and roots

Humans rely on a more diverse source of foods, especially more meat

Metabolic demands of a larger brain

Human brain volume much larger than that of human-chimp ancestor

Metabolic demands scale linearly with volume

Modifications in metabolism necessary to compensate for larger brains

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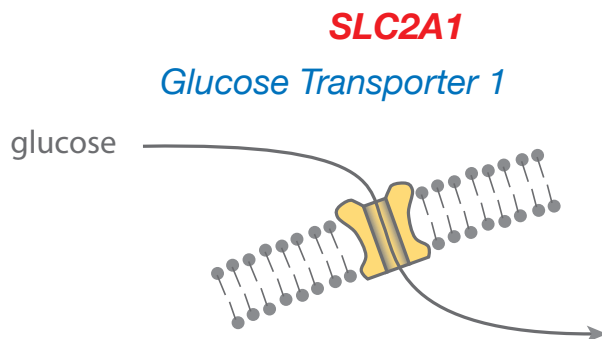
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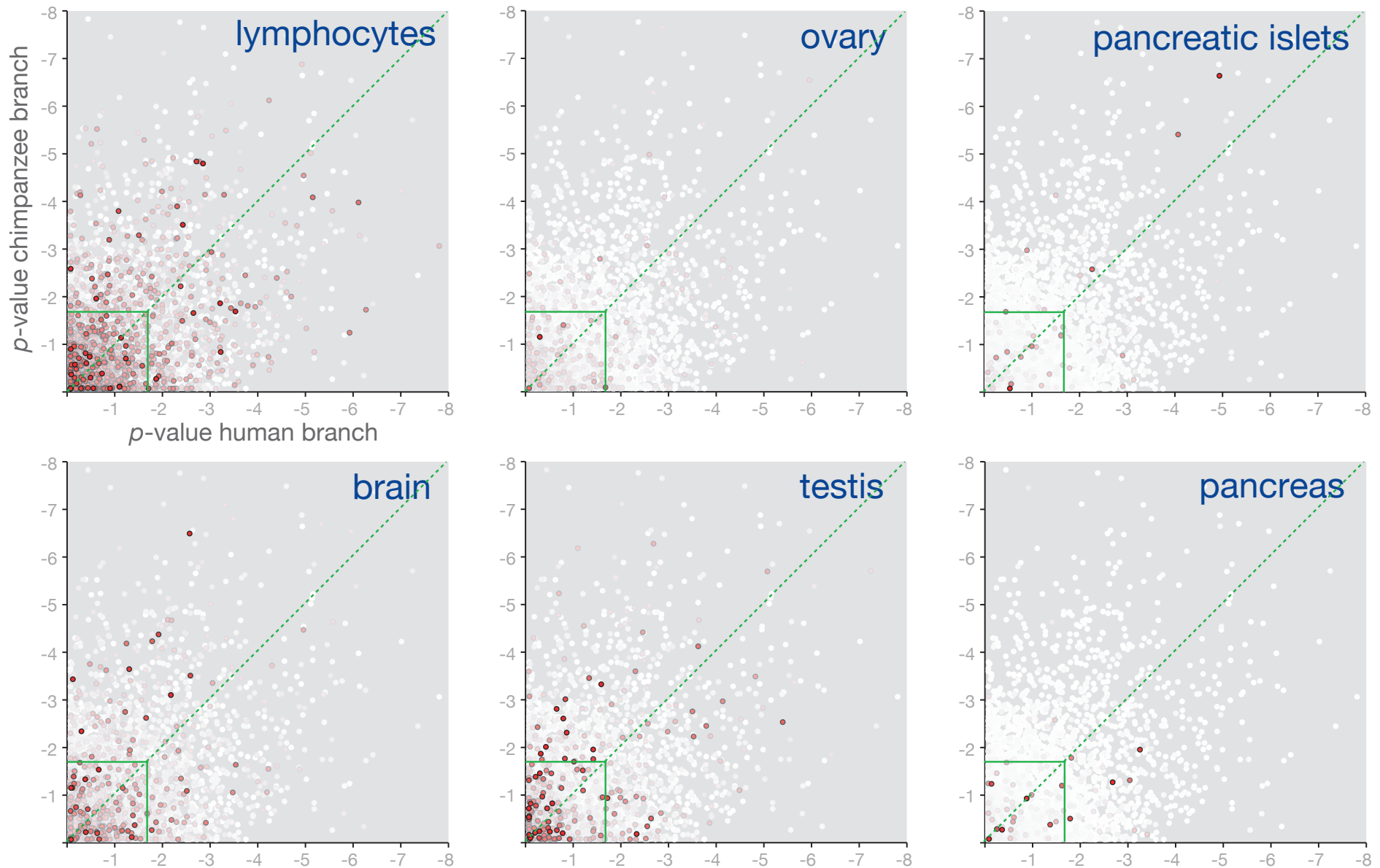


Major glucose transporter across blood-brain barrier

Association: noninsulin-dependent diabetes mellitus

Hemizygous: acquired microcephaly, cognitive impairments

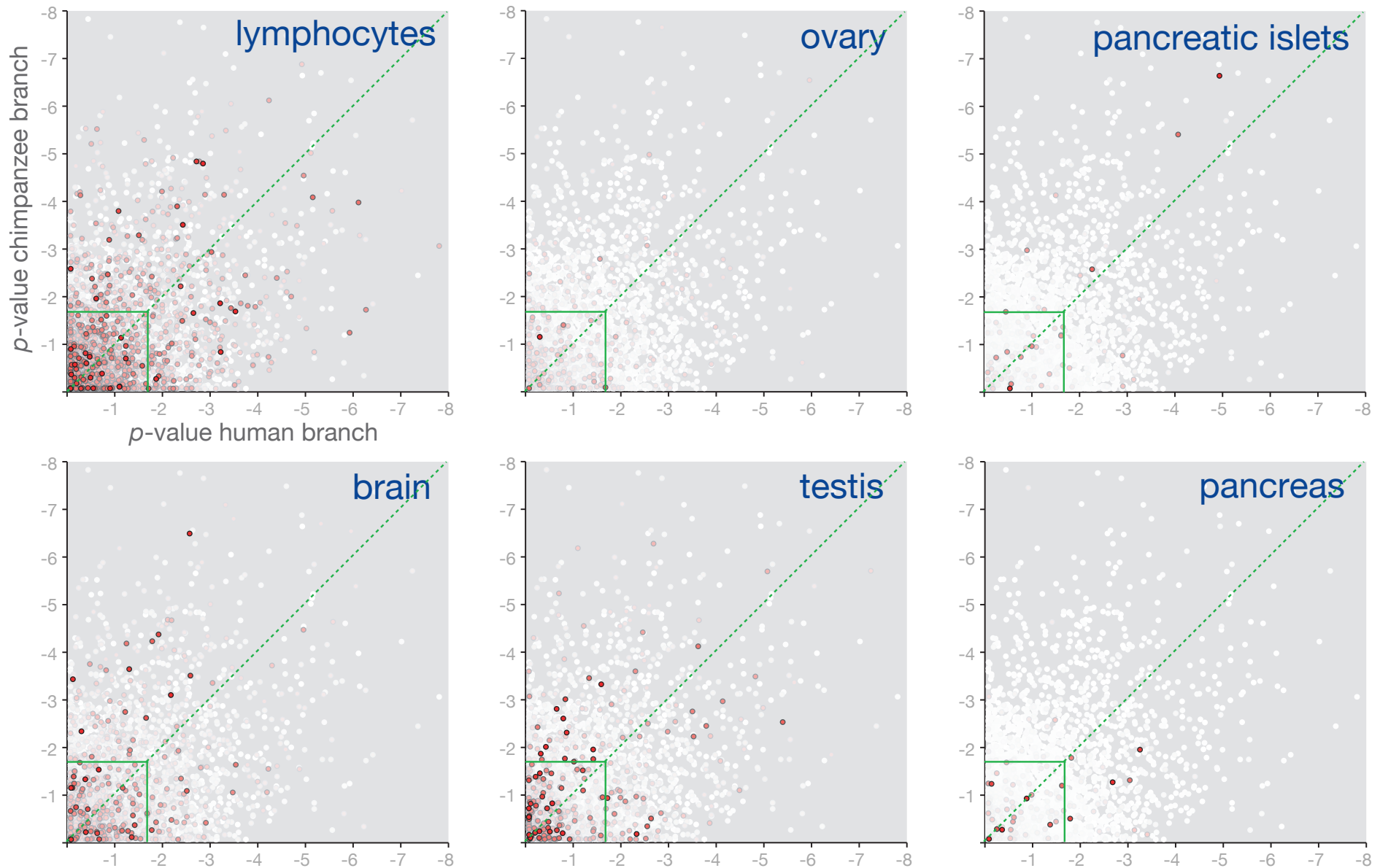
Branch-specific positive selection and expression specificity



Expression of 6,316 genes in 73 tissues from Novartis Human Gene Expression Atlas.

Expression specificity: *unique*  *ubiquitous*

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Why selection on the chimpanzee branch?



Chimpanzee art, painted by Congo in 1957

King and Wilson's proposal

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King, M-C and A Wilson (1975) *Science* 188:107

More positive selection on 5' flanking than coding regions

<i>partition</i>	<i>source</i>	<i>proportion</i>	<i>counts</i>
5' flanking	our analysis	0.0041	26/6316
coding	Clark et al.	0.0000	0/7645
5' flanking	our analysis	0.0140	88/6316
coding	Nielsen et al.	0.0019	21/11159

Counts based on q -values < 0.05

Sources: Clark et al. (2003) *Science* 302:1960; Nielsen et al. (2005) *PLoS Biology* 3:976.

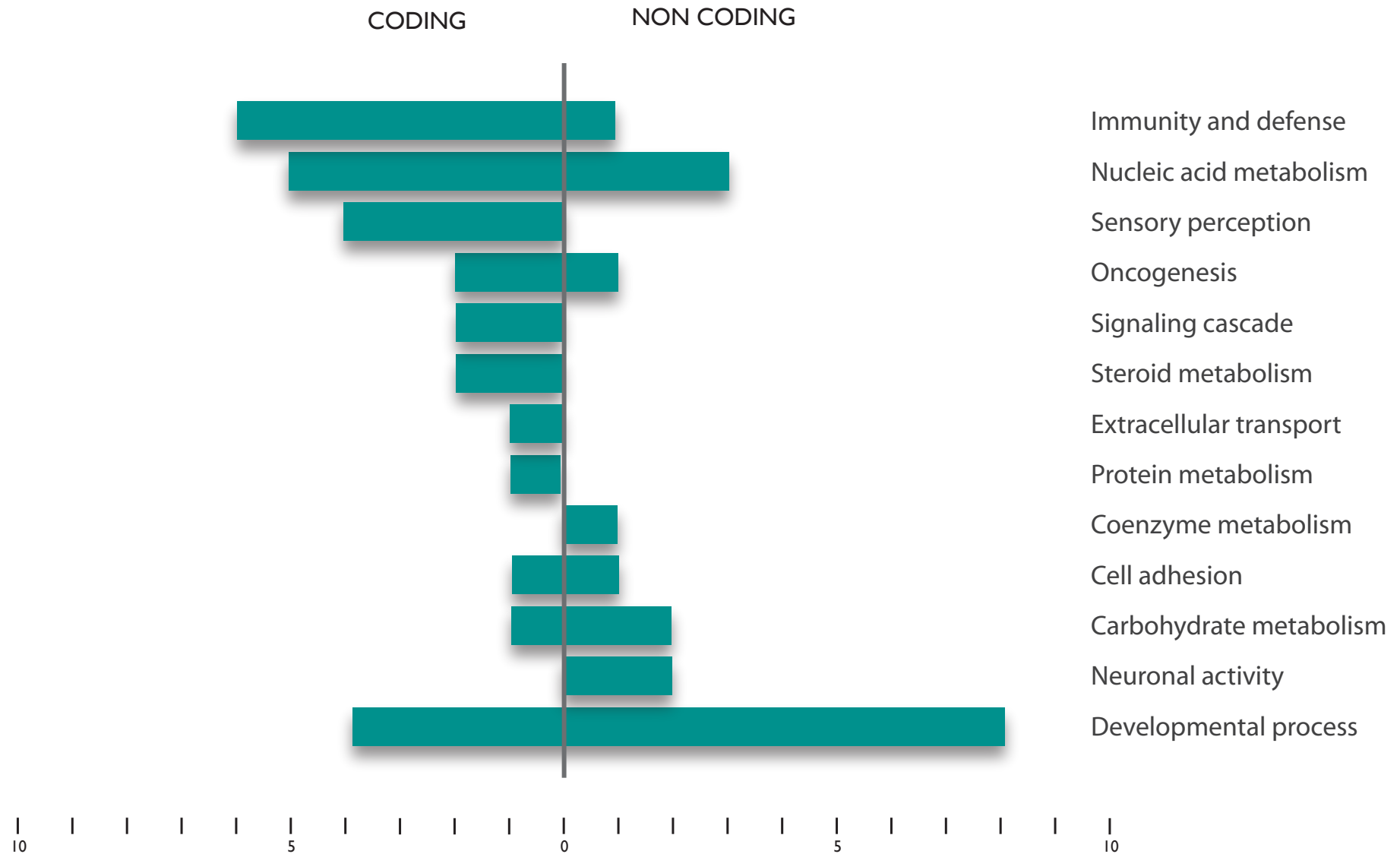
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Sources: Clark et al. (2003) *Science* 302:1960; Nielsen et al. (2005) *PLoS Biology* 3:976.

Qualitative differences in selection on coding/regulatory regions



Meta-analysis of published genome-wide scans for selection: three coding region and two non-coding region.
All Panther Biological Process categories with meta p-value < 0.05, sum of log-weighted meta p-values

Part 1 summary

- Lots of positive selection on regulatory sequences
- Neural, metabolic functions prominently affected
- Many new candidate genes for further study

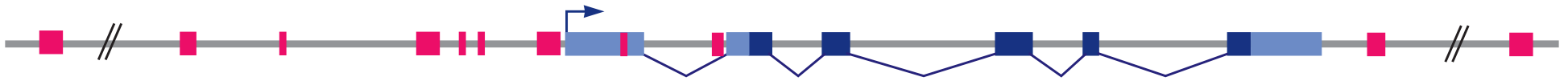
screening for more candidate genes

testing for functional differences

connecting the dots: 3 case studies

Studying *cis*-regulatory evolution

the bad news: *cis*-regulatory sequences are cryptic and spatially unbounded

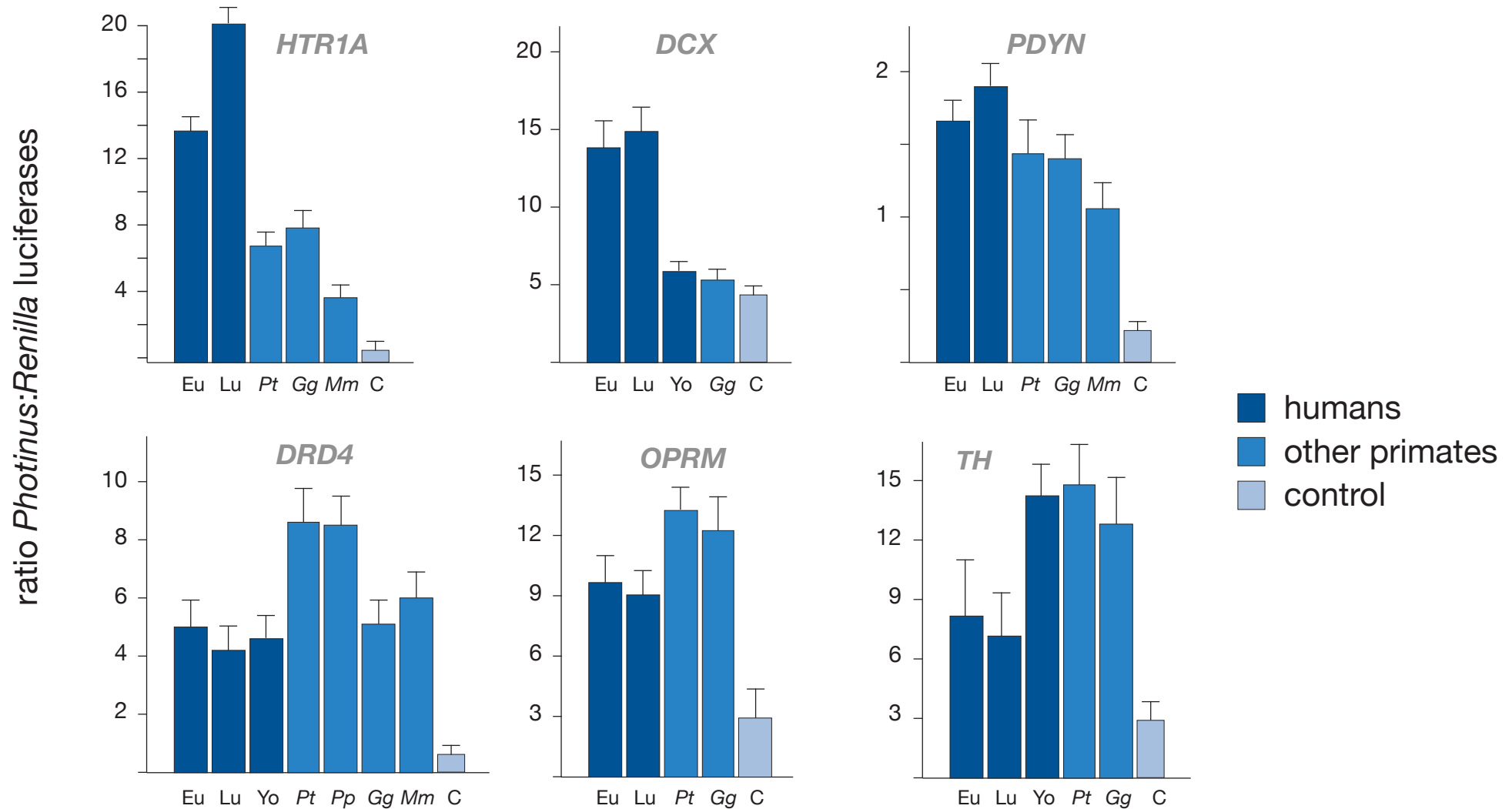


the good news: functional tests are easy and uniformly applicable

Testing for differences in gene expression

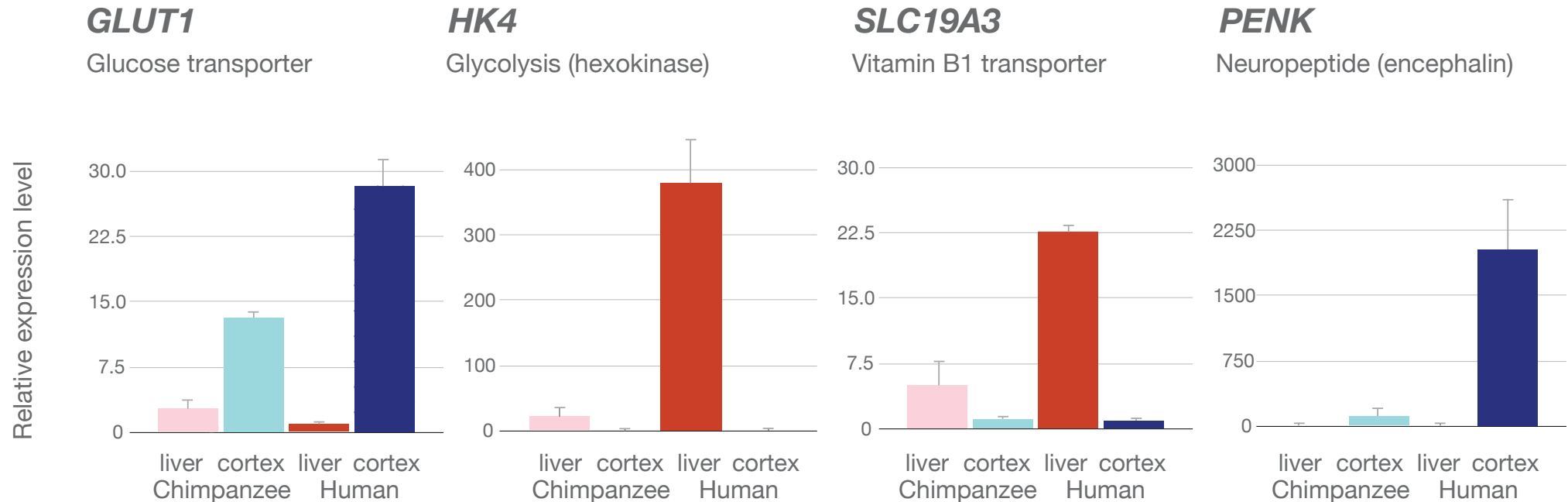
method	context	scale	genetics?	sensitivity	precision
reporter gene assay	<i>in vitro</i>	1-10s	yes	high	good
quantitative PCR	<i>in vivo</i>	1-100s	no/yes	high	good
pyrosequencing	<i>in vivo</i>	1-100s	yes	high	good
microarrays	<i>in vivo</i>	10,000s	no	low	fair
SAGE profiling	<i>in vivo</i>	10,000s	no	high	good

Expression differences by reporter gene assays



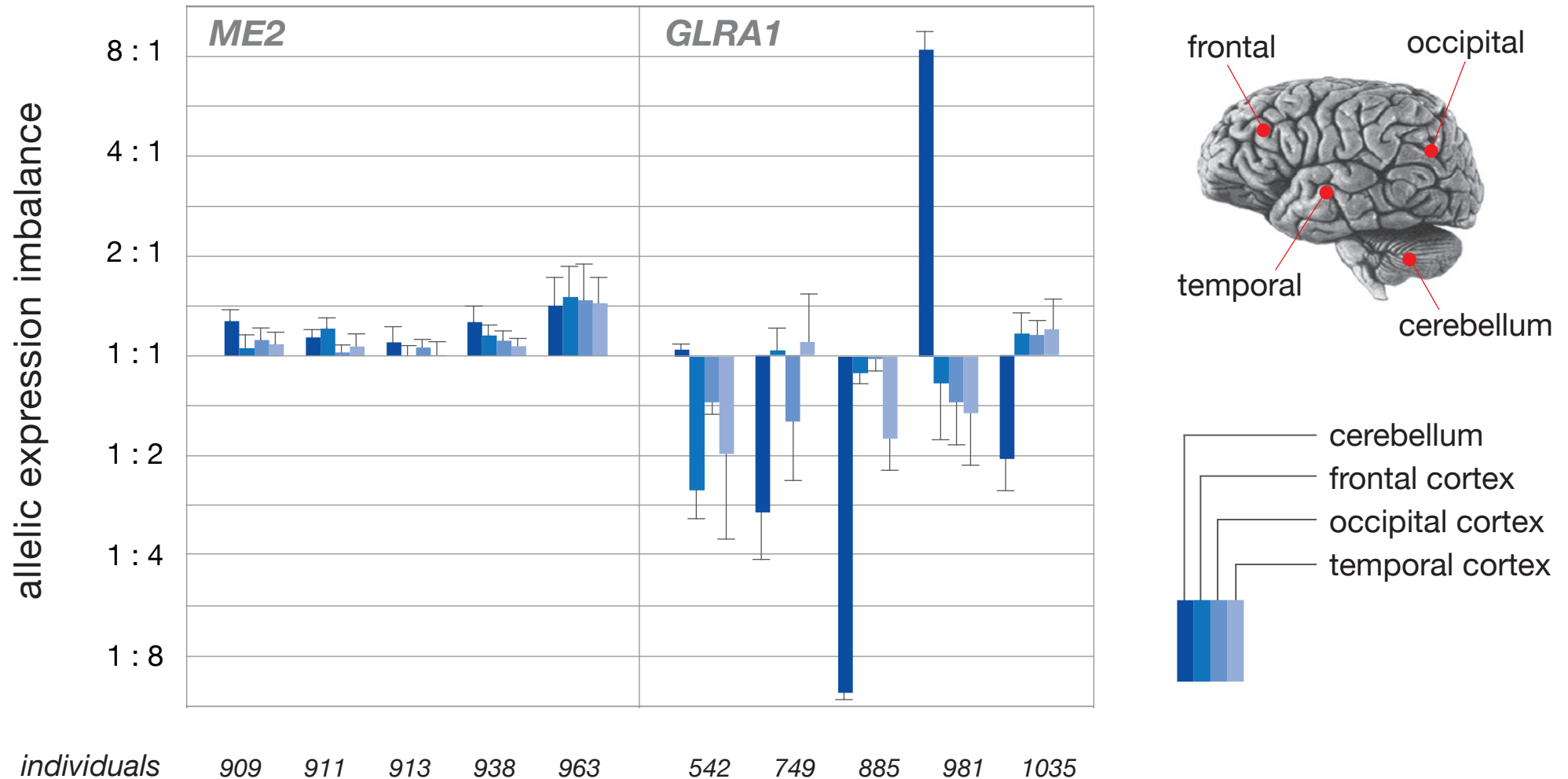
Six brain-expressed genes whose 5' flanking regions show strong signals of positive selection. Dual Luciferase Assay in IMR-32 or SH-SY5Y cells; bars are means of 8 technical replicates.

Expression differences between species by qPCR



Four genes of interest: 3 with strong signatures of positive selection, 4th is a candidate gene.
Quantitative PCR; values are means of 4 individuals and 6 technical replicates.
Error bars are standard deviations on all 24 measurements for each tissue x species.

Expression differences between alleles in the same individual



Two brain-expressed genes whose 5' flanking regions show strong signals of positive selection. Allelic imbalance measured by pyrosequencing; bars are means of 8 technical replicates.

Part 2 summary

- Same functional tests can be applied to almost any gene
- Tests for functional changes in regulation are informative
- Positively-selected genes often show functional changes

screening for more candidate genes

testing for functional differences

connecting the dots: 3 case studies

LCT

Encodes lactase-phlorizin hydrolase

Dual-function enzyme

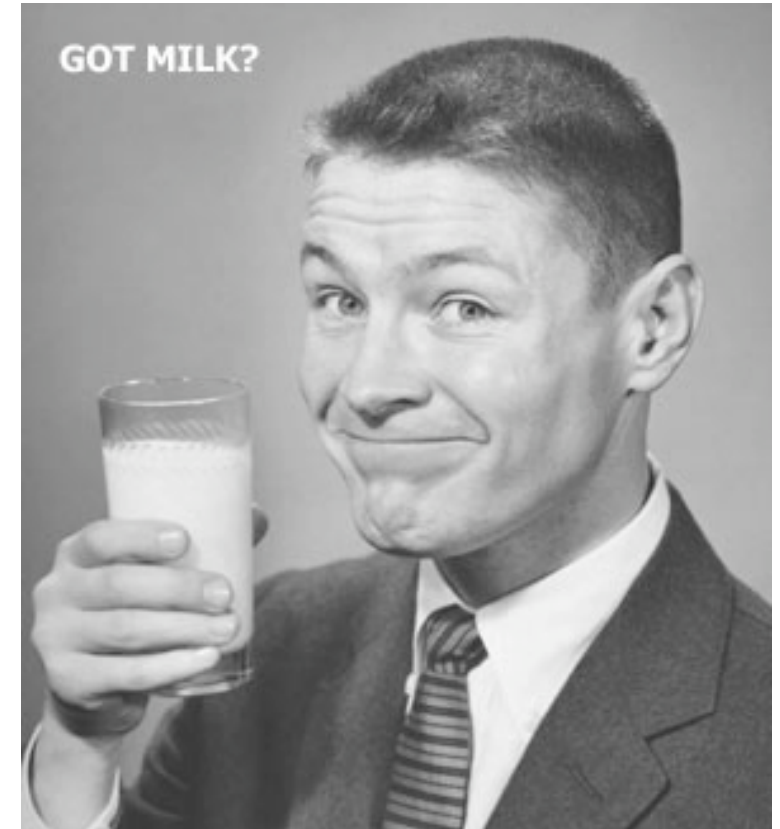
Digests lactose

Responsible for lactase persistence

Ability to metabolize lactose as adults

Evolved very recently (< 20,000 years)

Associated with pastoralism



An enzyme associated with a uniquely human metabolic function

Lactase persistence: genotype and phenotype

Phenotype

- LP
- LIP
- LNP

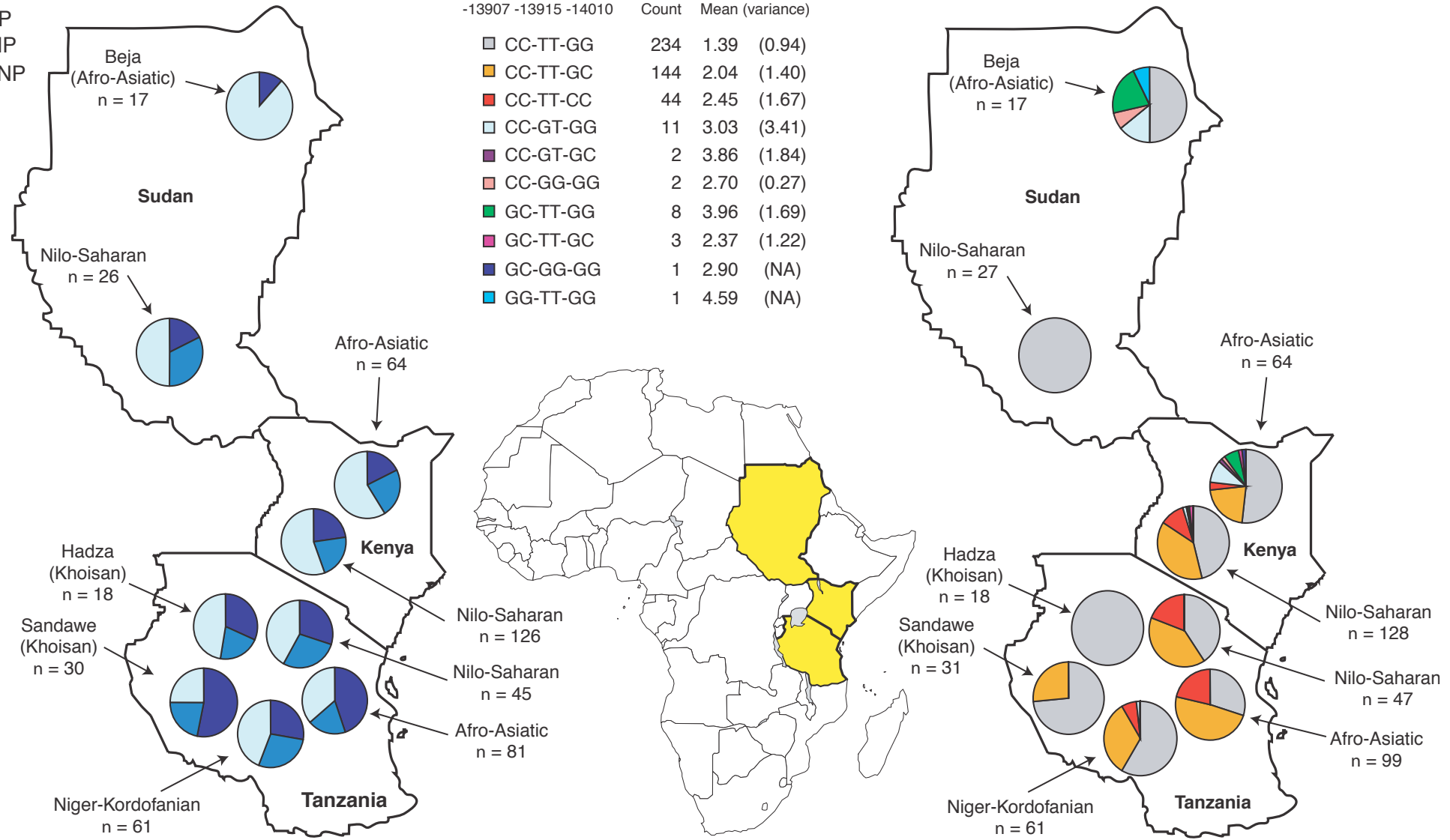
Genotype

-13907 -13915 -14010

Genotype	Count	Mean (variance)
CC-TT-GG	234	1.39 (0.94)
CC-TT-GC	144	2.04 (1.40)
CC-TT-CC	44	2.45 (1.67)
CC-GT-GG	11	3.03 (3.41)
CC-GT-GC	2	3.86 (1.84)
CC-GG-GG	2	2.70 (0.27)
GC-TT-GG	8	3.96 (1.69)
GC-TT-GC	3	2.37 (1.22)
GC-GG-GG	1	2.90 (NA)
GG-TT-GG	1	4.59 (NA)

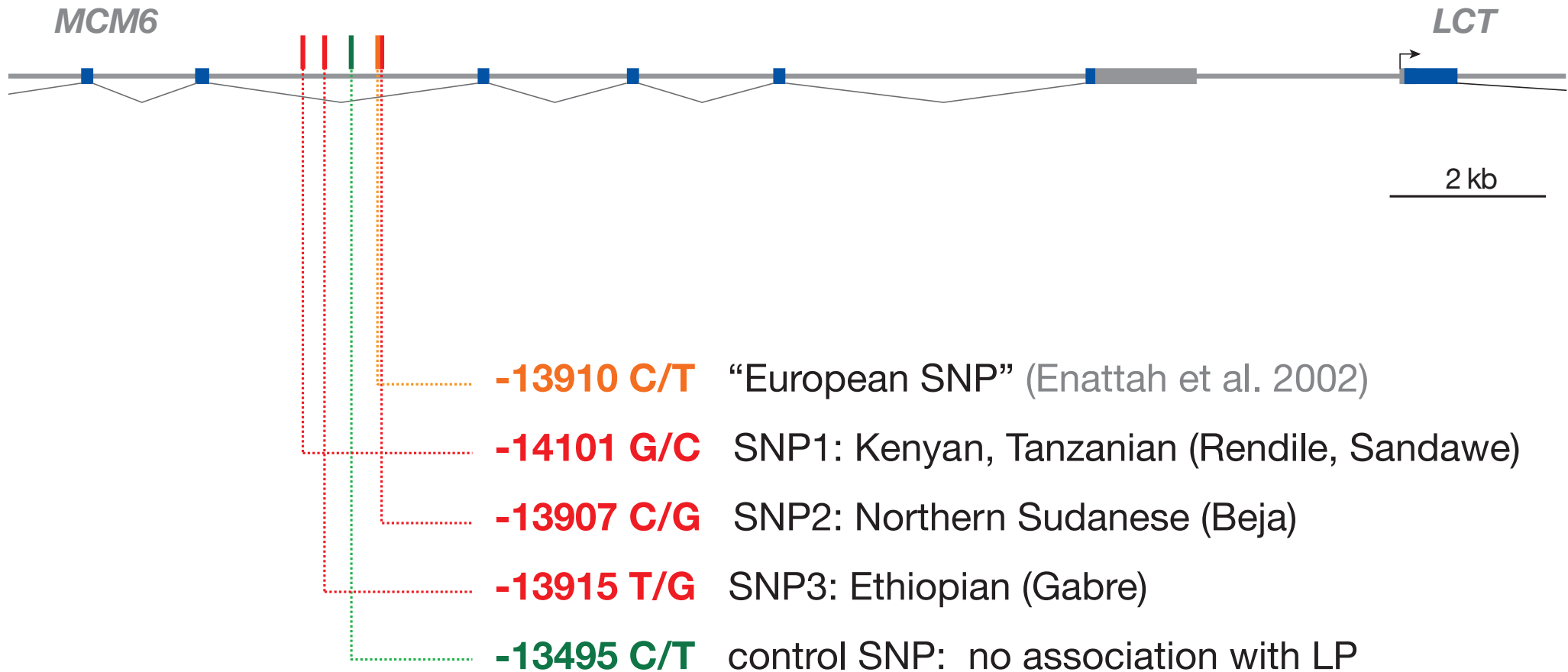
Phenotype

Count Mean (variance)

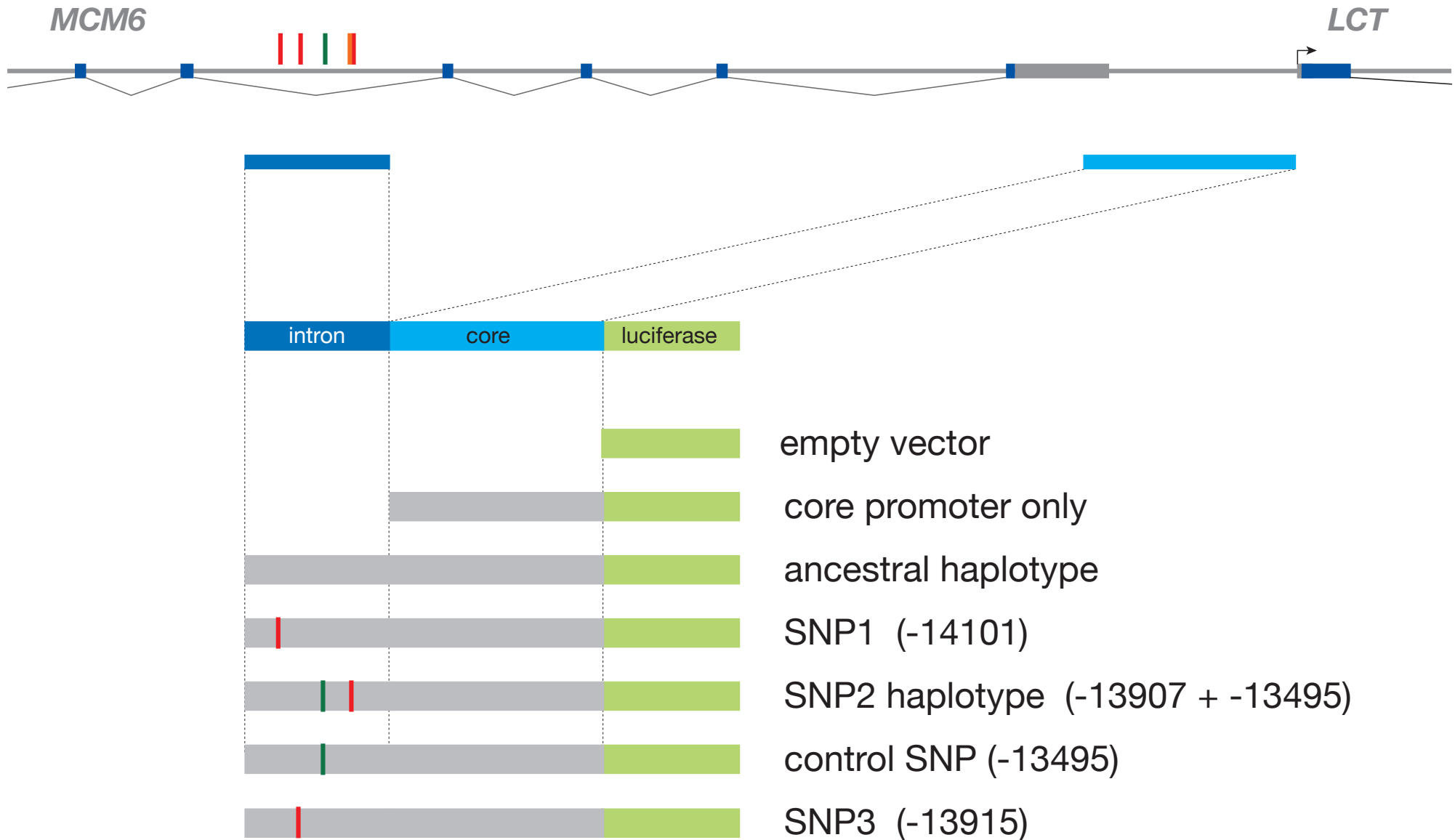


Collaboration with: Sarah Tishkoff, Jonathan Pritchard, Panos Deloukas

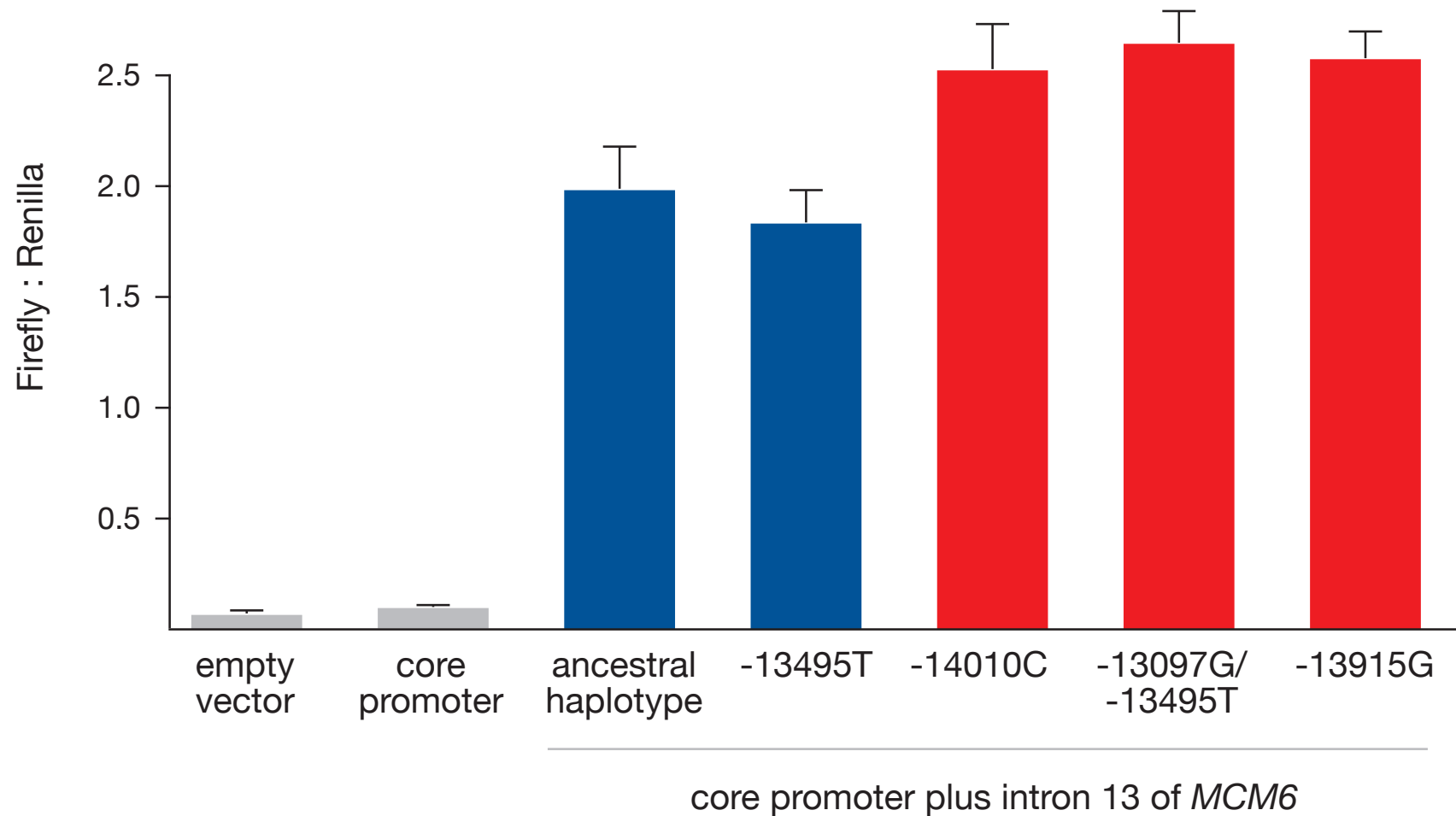
Genetic basis: implicated SNPs



Constructs for testing implicated SNPs

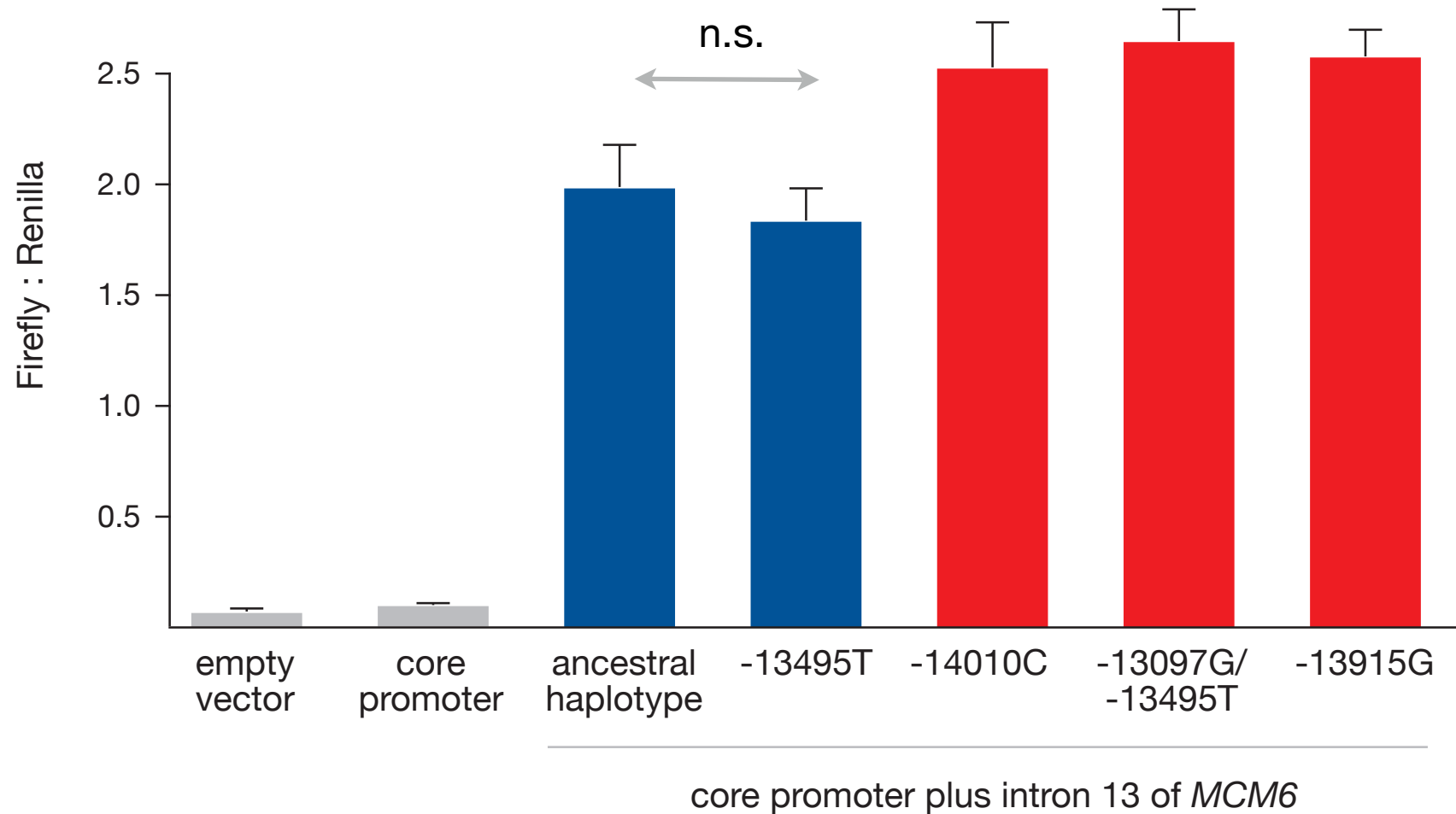


All three implicated SNPs elevate transcription



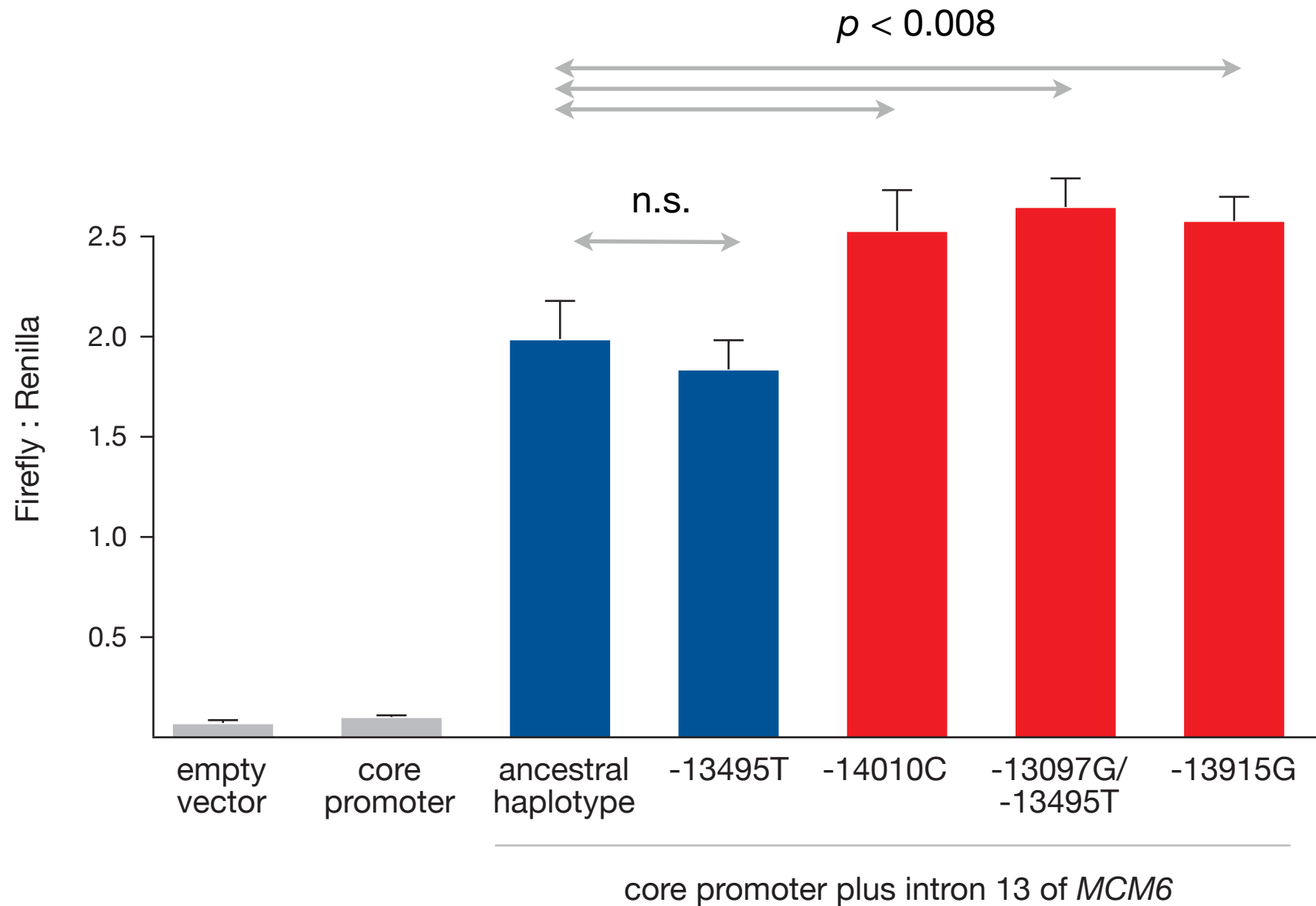
Dual-Luciferase assay. Bars are means of 12 technical replicates.

All three implicated SNPs elevate transcription



Dual-Luciferase assay. Bars are means of 12 technical replicates.

All three implicated SNPs elevate transcription



Dual-Luciferase assay. Bars are means of 12 technical replicates.

The ecological agent



Hans-Peter Harmsen

DARC

Encodes chemokine receptor Duffy

Gene formerly known as *FY*

Signaling within the immune system

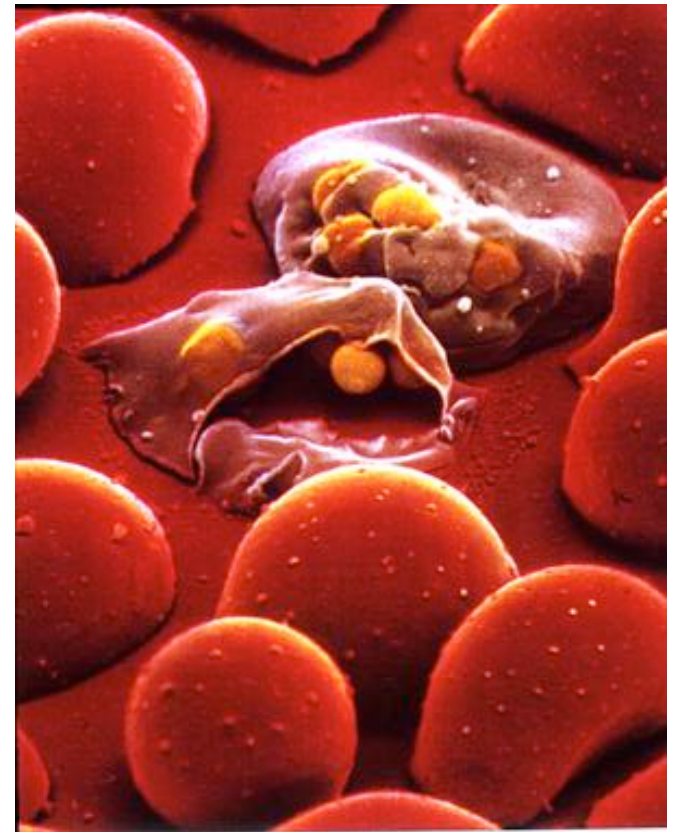
Resistance to *Plasmodium vivax* infection

SNP -32 abolishes erythrocyte expression

Monogenic trait, semi-dominant

Allele fixed in regions of malarial endemism

Clear signature of natural selection



University of Florida School of Medicine

A dramatic example of adaptation in cis-regulatory function

Savannah baboons in the Amboseli basin, Kenya

Project of Stuart Altmann, Jean Altmann, and Susan Alberts

Continuous field observation and detailed data from 1971-present

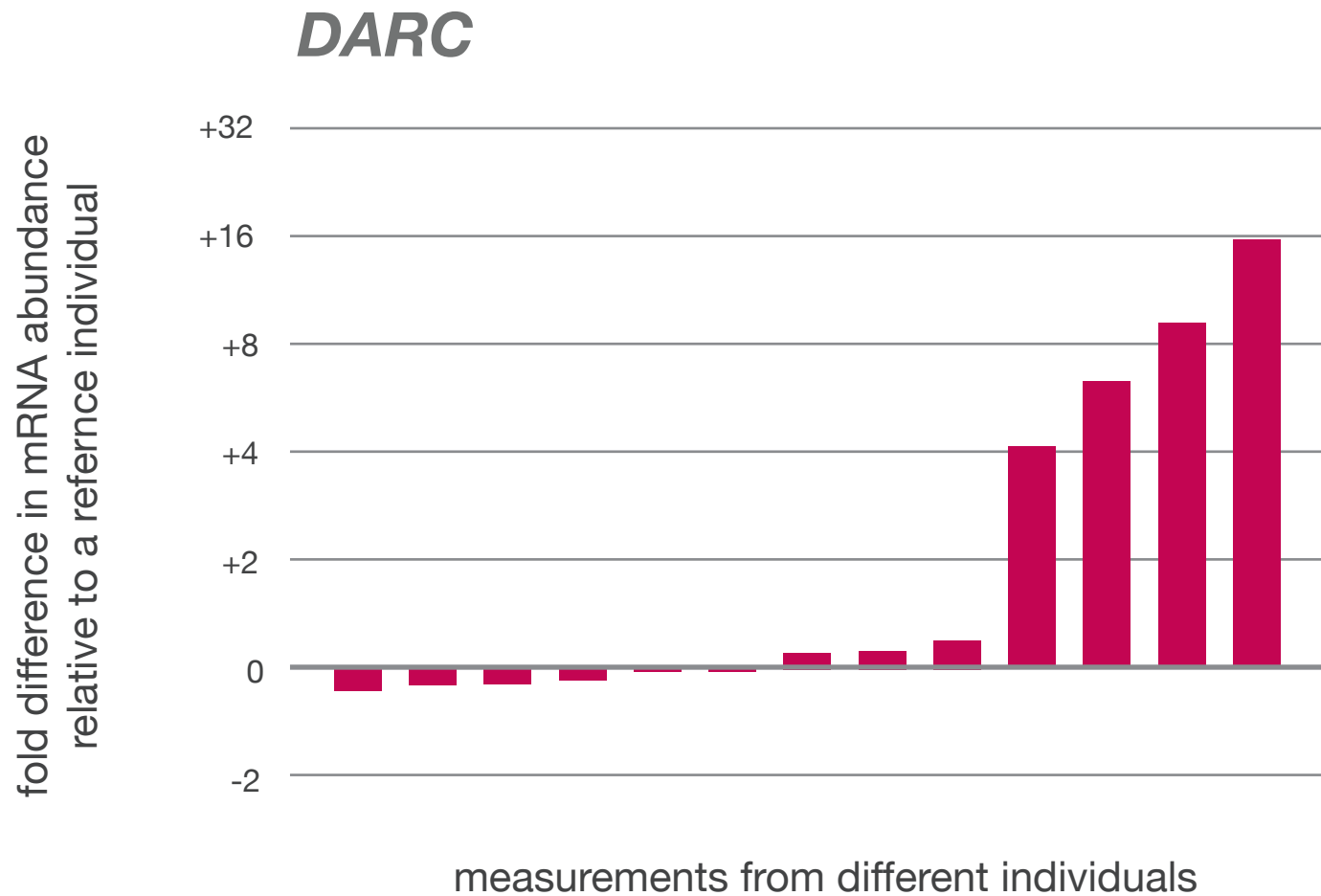
Data from >1000 individuals, many known pedigrees (3-5 generations)

Genomic DNA from >600 individuals

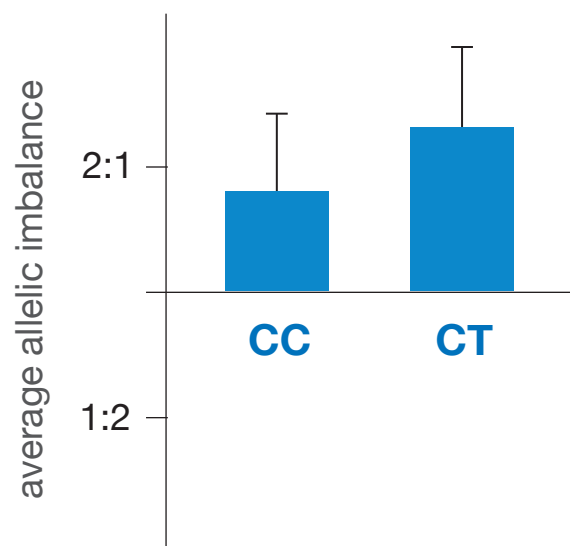
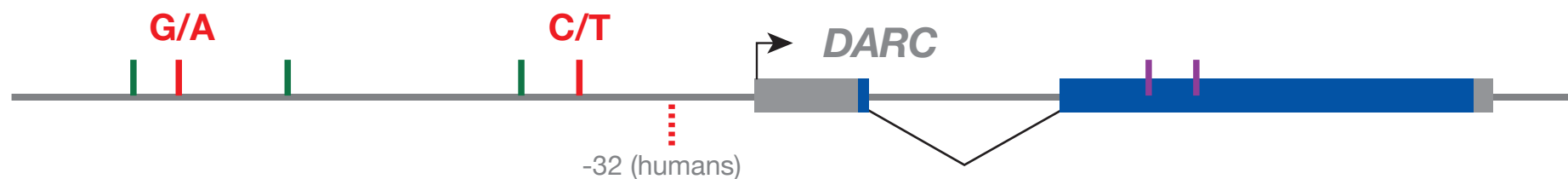
cDNA from peripheral blood and skin biopsies from >80 individuals



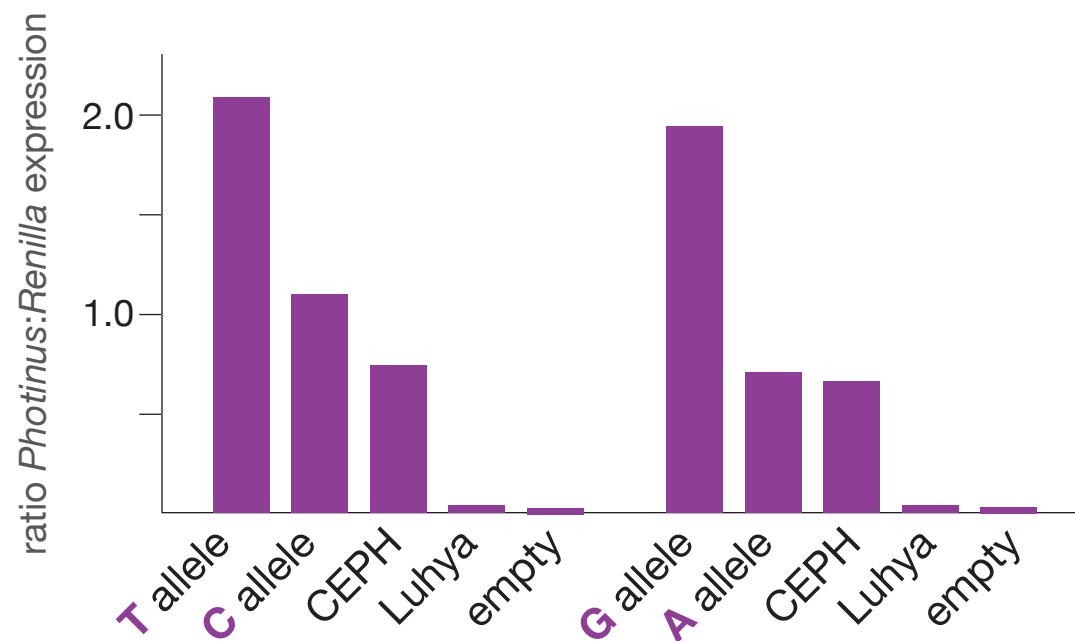
In vivo differences in transcript abundance in wild individuals



Two 5' SNPs near *DARC* influence expression levels

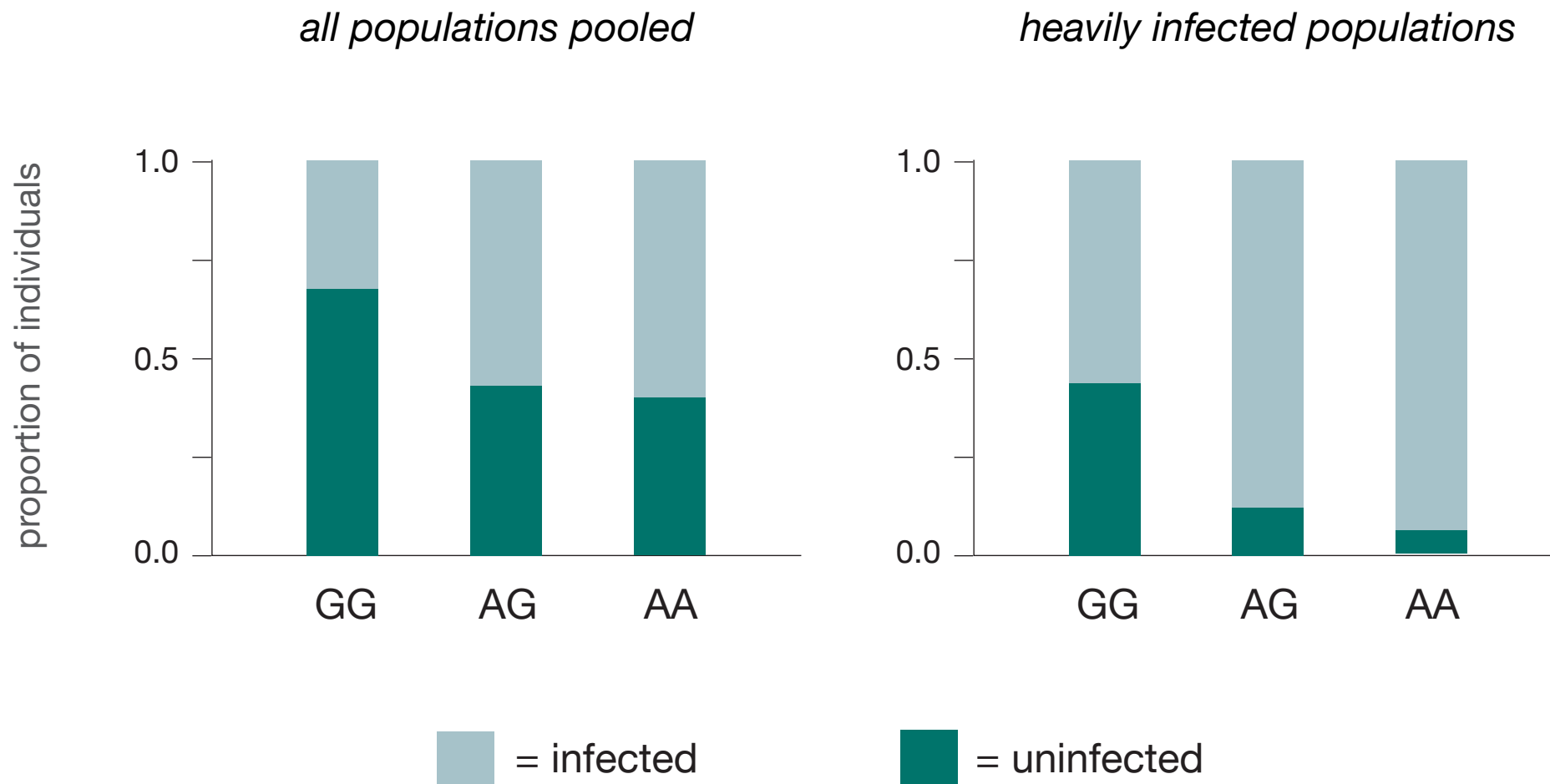


Genotype of proximal SNP predicts *in vivo* allelic imbalance ($p < 0.047$)



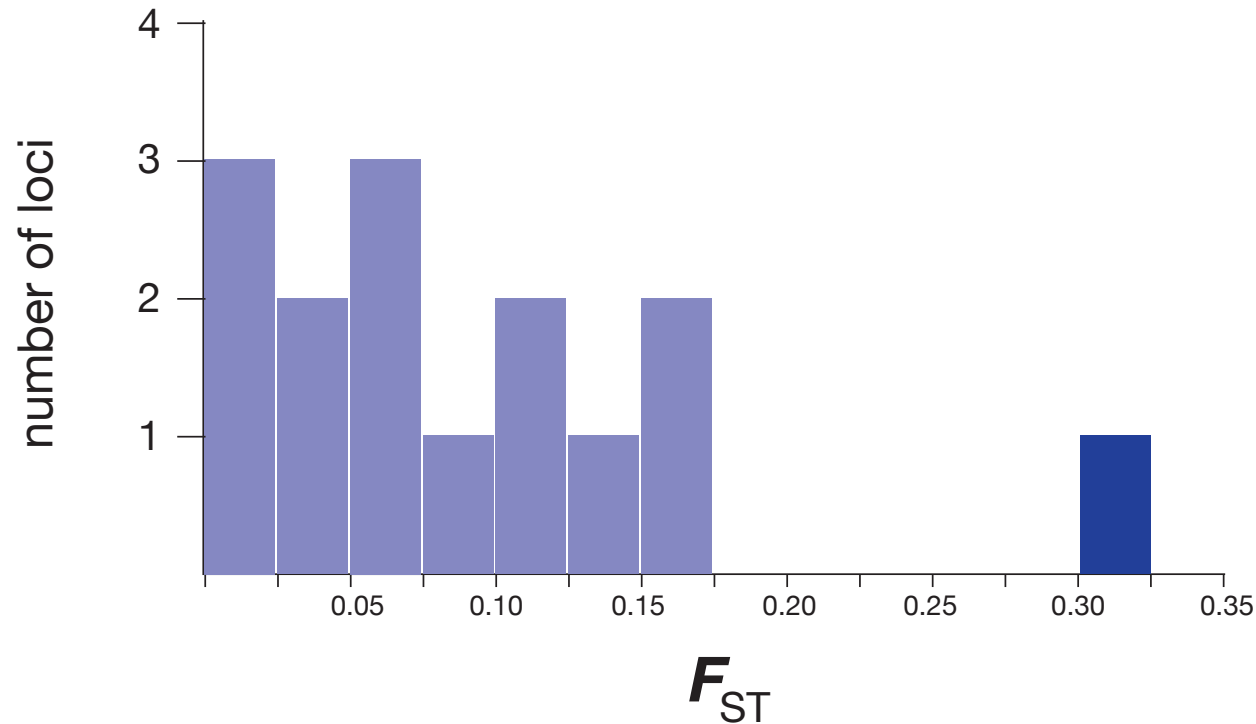
Relative expression levels, functional assays *in vitro* in HEL cells (all between construct differences significant)

Distal SNP genotype correlated with *Hepatocystis* infection



Natural infections of baboons by: *Plasmodium knowlesi* and closely related haematoprotezoan parasites within the in genera *Babesia*, *Entoployploides*, and *Hepatocystis*.

Signatures of selection on the 5' flanking region of *DARC*



Significant differentiation ($p < 0.001$) among three wild baboon populations: Amboseli (Kenya), Masai Mara (Kenya), and Mikumi (Tanzania), suggesting local directional selection.

Tajima's D also significantly elevated ($D = 1.755$, $p < 0.024$), suggesting balancing selection.

DRD4

Encodes a dopamine receptor

Metabotropic G protein-coupled receptor

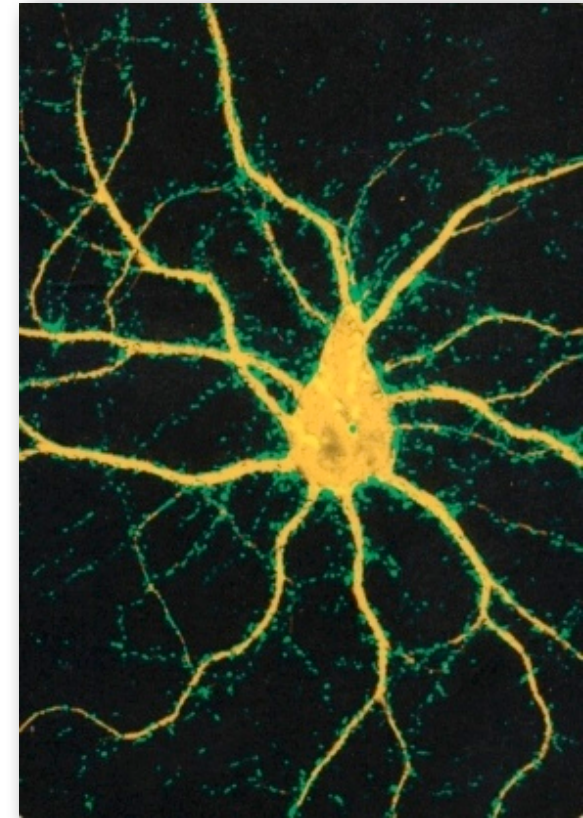
Inhibitory to post-synaptic neuron

Genetically associated with personality traits

Novelty-seeking, impulsiveness, risk-taking

Attention-deficit hyperactivity disorder

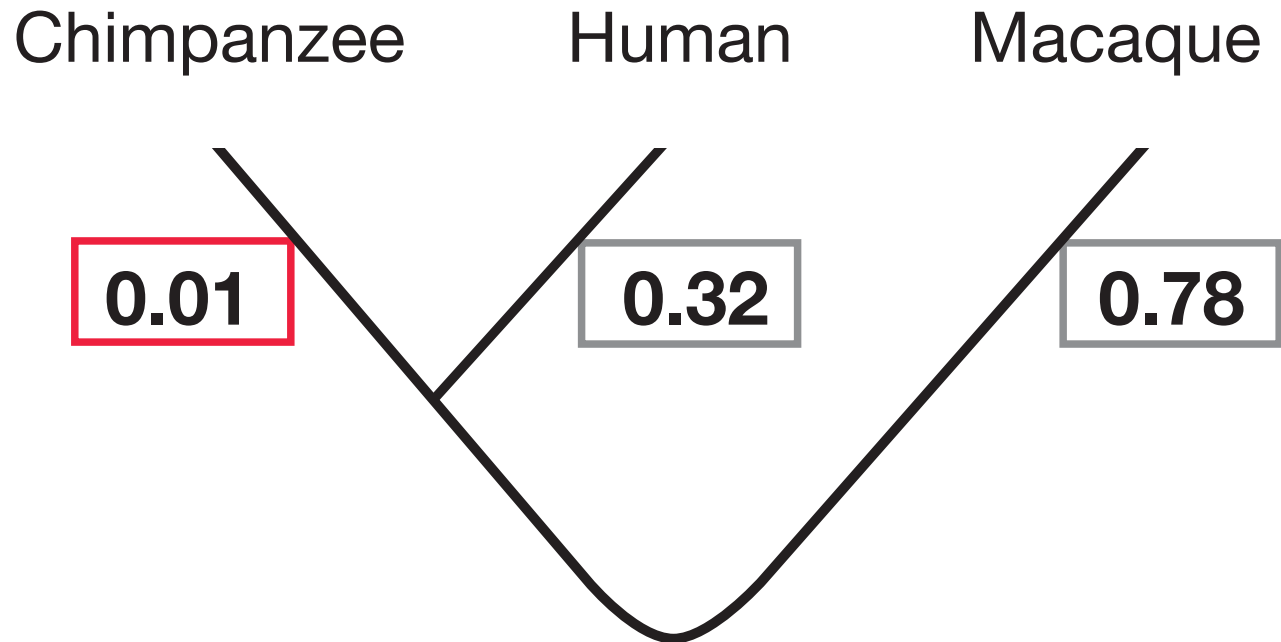
Schizophrenia



Peter Vanhoenacker

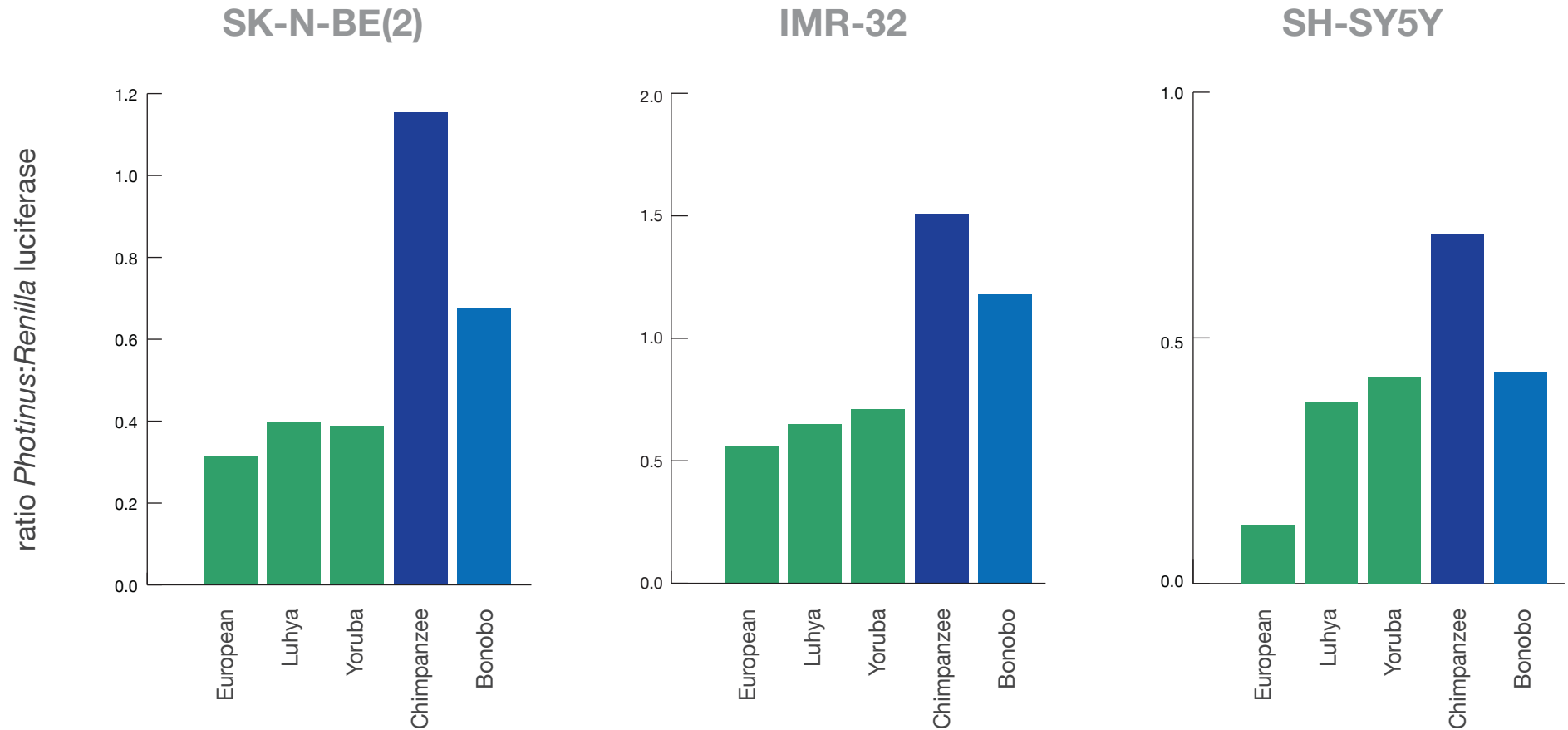
A protein associated with variation in cognition in humans

Branch-specific positive selection on *DRD4* 5' flanking region



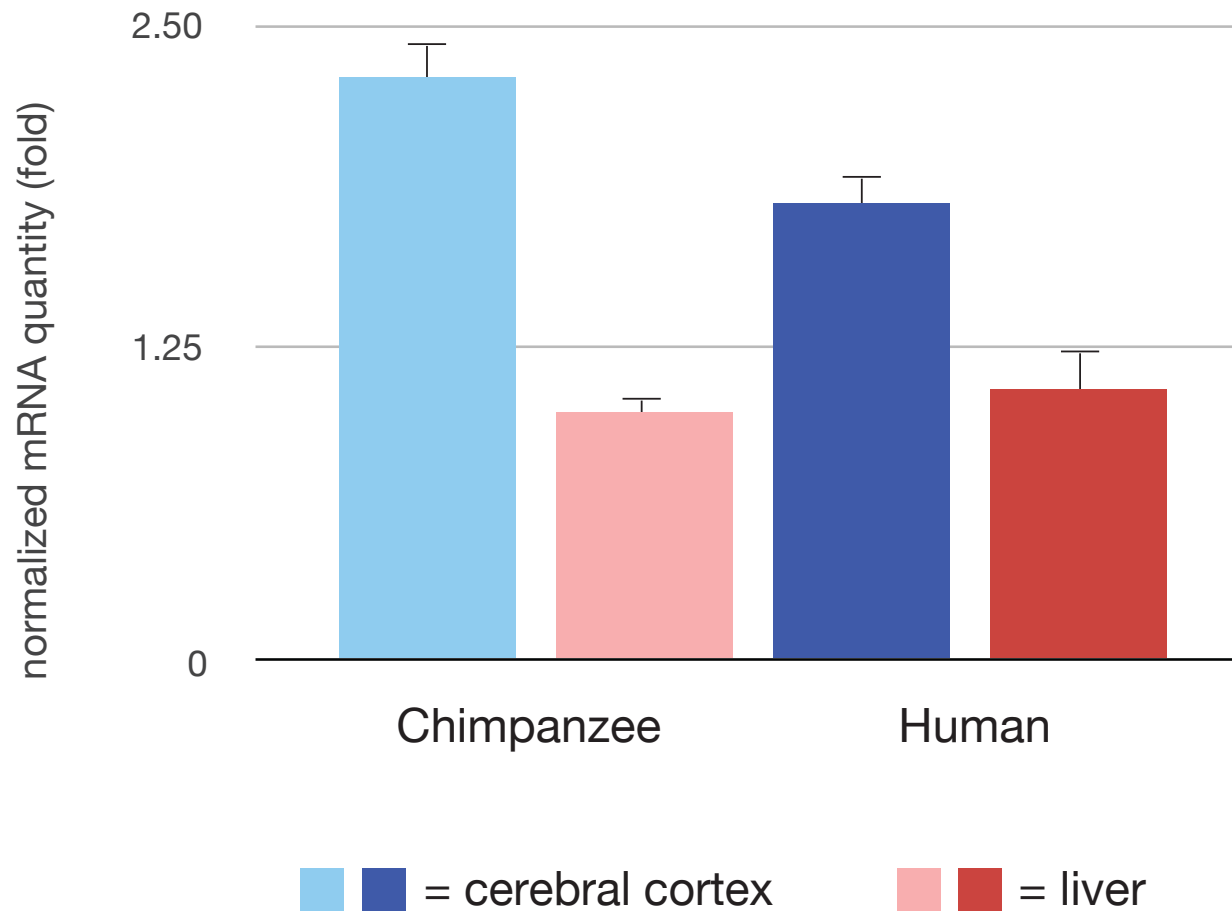
p-values; random-effects likelihood test for selection on 5 kb of 5' flanking region.

Chimpanzee 5' flanking region drives higher expression



Dual-Luciferase assay. Tested in three different neuroblastoma cell lines.

Chimpanzees have higher *in vivo* *DRD4* expression in brain



Quantitative PCR on cDNA made from mRNA extracted from autopsied tissue.
Bars are means of six technical replicates.

What traits are affected?



Bonobo

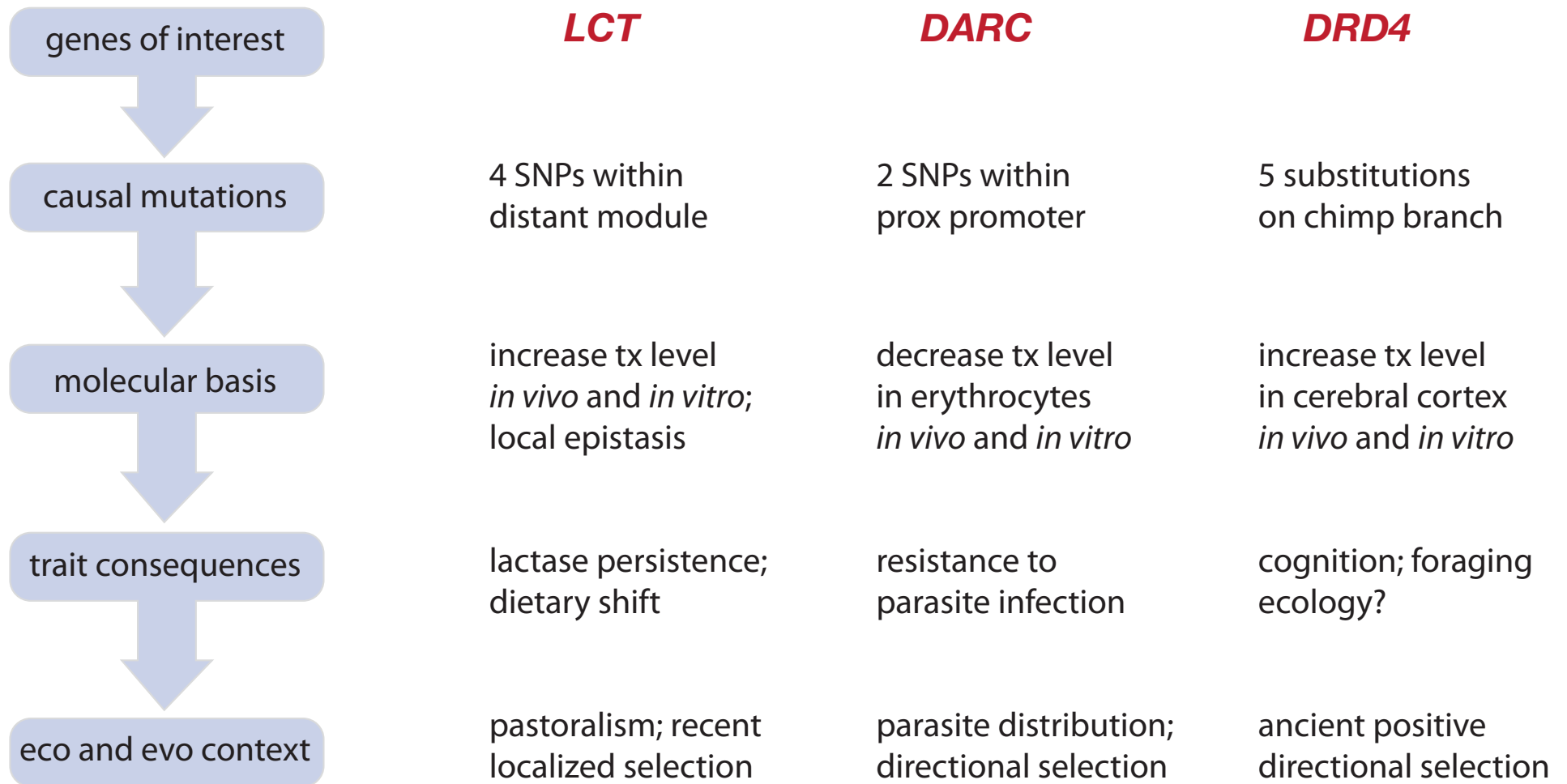
food-rich, less seasonal habitat
abundant, nutritious back-up food
casual foragers, indifferent to food
risk-averse on standard psych profile



Chimpanzee

seasonal habitats
variable, nutrient poor back-up foods
active foragers, fixated on food
risk-tolerant on same profile

Part 3 summary: connecting the dots



Thanks to many people . . .

Computational

Ralph Haygood
Olivier Fédrigo
Jovana Pavisc
Brian Hanson
Ken Yokoyama

Humans

Courtney Babbitt
Matt Rockman
Jesse Silverman
Lisa Warner
William Nielsen
Tonya Severson
Adam Pfefferle
Shauna Morrow

Baboons

Jenny Tung
Alex Primus
Dagan Loisel
Andrew Bouley
Tonya Severson

Help with programming

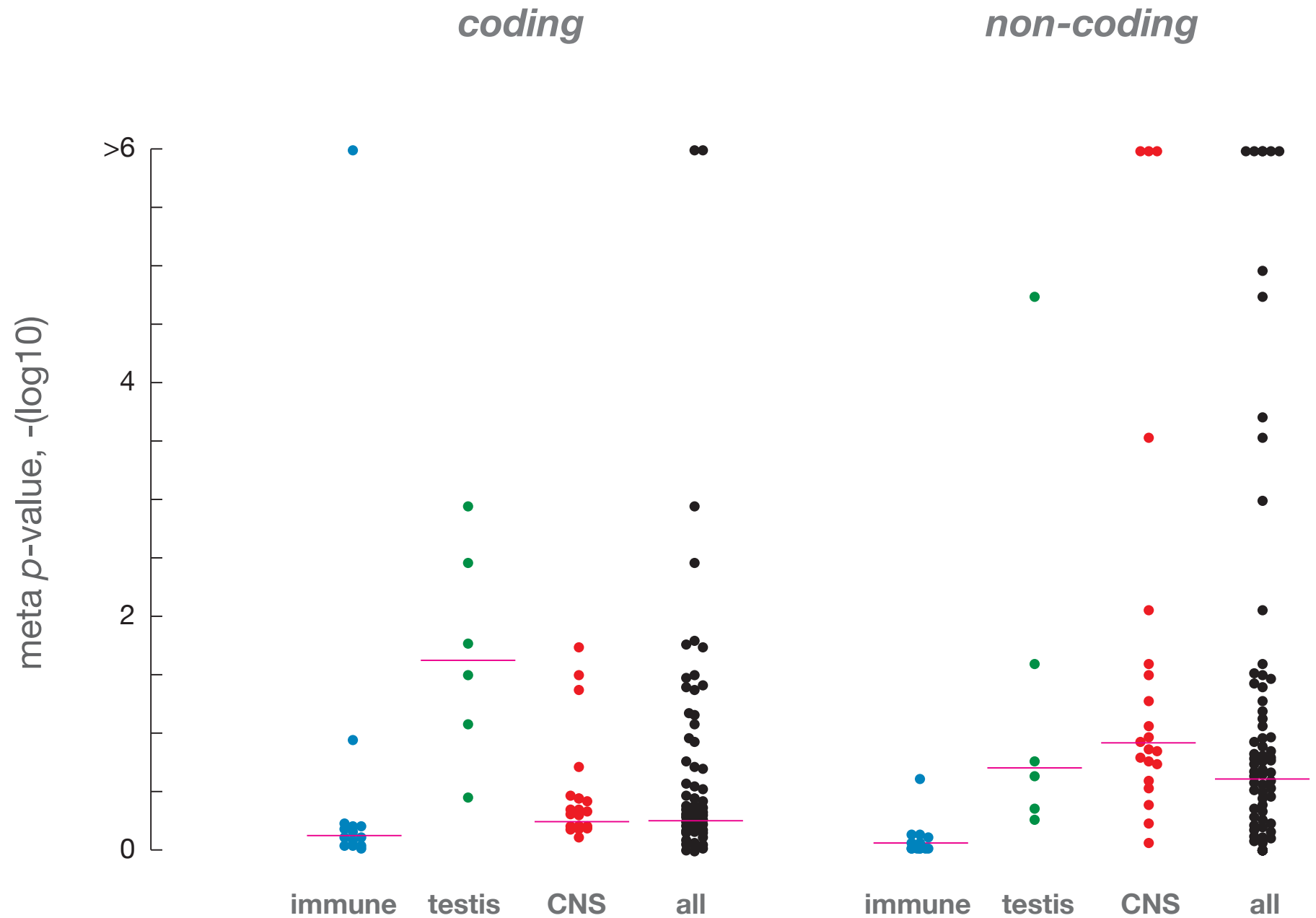
Jim Kent (UCSC)
Sergey Kosakovsky-Pond (UCSD)
Webb Miller (Penn)

Collaborators

Sarah Tishkoff (U MD)
Susan Alberts (Duke)
Jean Altmann (Princeton)



Qualitative differences in selection on coding/regulatory regions



Understanding the evolution of gene expression

Genetic basis

Genes involved

Causal mutations

Interactions among genes

insights

Understanding the evolution of gene expression

Genetic basis

Genes involved
Causal mutations
Interactions among genes

Functional basis

Biochemical changes
Molecular interactions
Cell/development impact

insights

A Venn diagram with two overlapping light blue circles. The left circle is labeled 'Genetic basis' and contains the text 'Genes involved', 'Causal mutations', and 'Interactions among genes'. The right circle is labeled 'Functional basis' and contains the text 'Biochemical changes', 'Molecular interactions', and 'Cell/development impact'. The intersection of the two circles is a darker blue shape labeled 'insights' in white text.

Understanding the evolution of gene expression

Genetic basis

Genes involved
Causal mutations
Interactions among genes

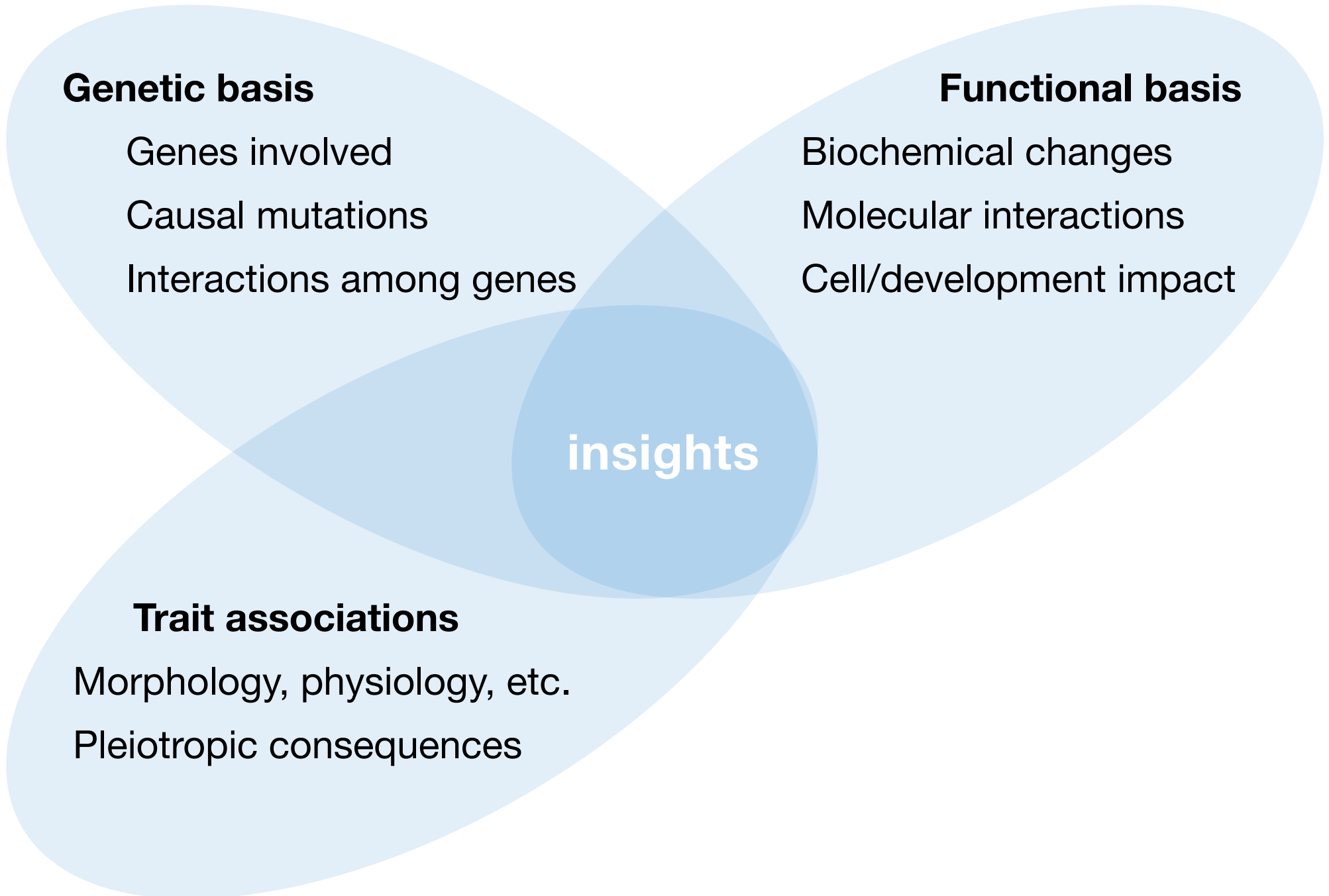
Functional basis

Biochemical changes
Molecular interactions
Cell/development impact

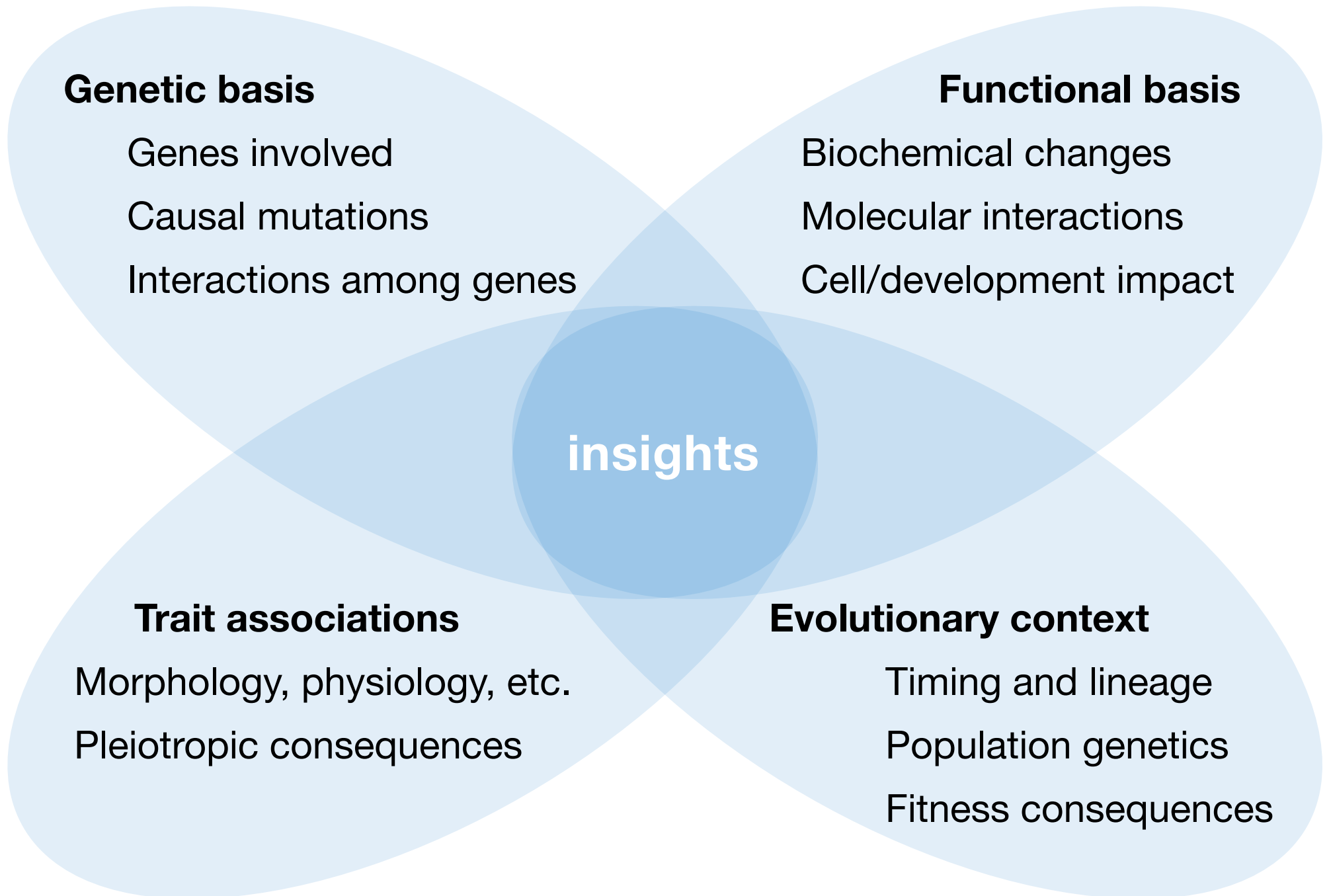
insights

Trait associations

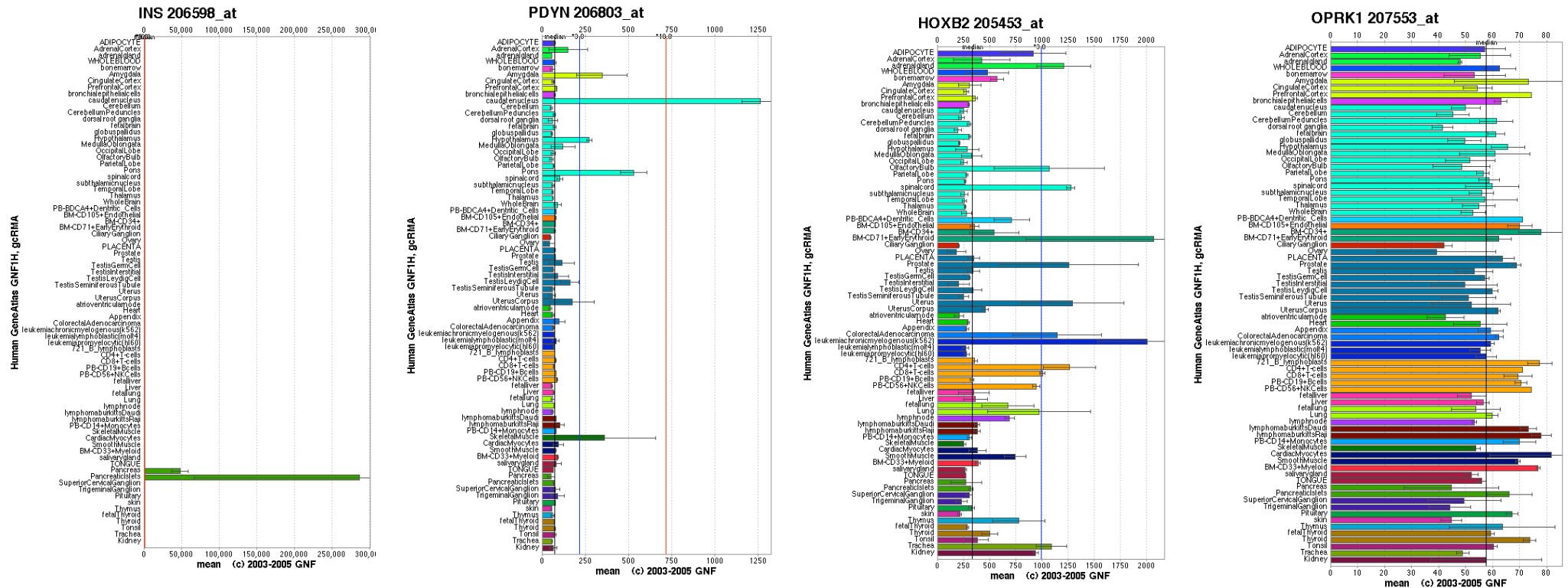
Morphology, physiology, etc.
Pleiotropic consequences



Understanding the evolution of gene expression



A metric for cell or tissue expression specificity



Data set: 73 non-cancerous tissues in the Novartis Gene Expression Atlas

Regard expression level of each tissue as a vector in 73-dimensional space

Specificity for each tissue = square of the cosine of the angle of the vector to that axis

Features

Independent of expression level

Can be defined for any gene in relation to any particular tissue